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Name of Document:	Compliance Status Report
Date of Document:	January 9, 2018
Site Name:	Fire Station 19, North Berkeley Lake Road Site
Site ID Number:	10844

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I certify that the information I am submitting is, to the best of my knowledge and belief, true, accurate, and complete.		Receipt Date (for EPD use only)
Signature: 0100444		
Name (printed):	J. Thomas Duffey, PG	
Date:	1/9/2017	
Organization:	CDM Smith	
Phone:	770-329-7143	
Email:	DuffeyJT@CDMSmith.com	

Department of Water Resources

684 Winder Highway • Lawrenceville, GA 30045-5012 678.376.6700 • www.gwinnetth2o.com

January 8, 2018

Mr. Tom Brodell Georgia Environmental Protection Division Land Protection Branch 2 Martin Luther King, Jr. Drive SE Suite 1456 East Atlanta, Georgia 30334

Subject: Compliance Status Report North Berkeley Lake Road Site (HSI No. 10844) Duluth, Gwinnett County, Georgia

Dear Mr. Brodell:

Gwinnett County is submitting the enclosed Compliance Status Report (CSR) prepared by CDM Smith for the Fire Station 19 property on the North Berkeley Lake Road Site. The CSR includes the required summary of findings, certification of compliance, and certification by a qualified groundwater scientist. Upon submitting this CSR, we have also initiated the public notification requirements. The notification is enclosed that will be published by the Gwinnett Daily Post within the required seven days.

We look forward to receiving your review comments and completing the remaining work to have the fire station delisted. If you have any questions, please do not hesitate to contact me directly at (678) 376-6953 or richard.schoeck@gwinnettcounty.com.

Sincerely,

GWINNETT COUNTY DEPARTMENT OF WATER RESOURCES

Richard Schoeck, P.E., PMP Division Director of Project Controls

Enclosures

cc: Tom Duffey, CDM Smith Forrest Fields, Gwinnett County J.C. Lan, Gwinnett County John Reichling, CDM Smith

PUBLIC NOTICE

The Georgia Environmental Protection Division, Department of Natural Resources, State of Georgia (EPD) has placed this site on the Hazardous Site Inventory pursuant to its authority under the Hazardous Site Response Act and Rules promulgated thereunder. As required by the Rules for Hazardous Site Response, the responsible party for this site was required to investigate the site and submit a compliance status report to EPD summarizing the results of that investigation. EPD is currently reviewing the compliance status report to determine if corrective action is needed for regulated substances that have been released at this site. Before EPD decides whether corrective action is needed, the public has the opportunity to review the compliance status report and provide comments to EPD about the report.

Site Information:

North Berkeley Lake Road Site, Gwinnett County, Duluth, Georgia 30096, HSI # 10844. HSI # 10844 currently consists of four (4) parcels held by separate owners. This public notice is being made on behalf of one property that is owned by Gwinnett County, as listed below:

Property Owner Information:

Gwinnett County Fire Station 16, 3275 North Berkeley Lake Road, Duluth, Georgia 30096. Parcel Identification Number R6267-028.

Written and oral contact can be made to the following:

EPD Contact Person:

Tom Brodell, 2 Martin Luther King Jr Drive, SE Suite 1056 Atlanta, GA 30334. Phone: 404-657-8600.

Gwinnett County Contact Person:

Richard Schoeck, Gwinnett County Department of Water Resources, 684 Winder Highway, Lawrenceville, Georgia, 30045-5012. Phone: 678-376-6700.

A copy of the Compliance Status Report is available for review and copy at the Gwinnett County Public Library, Peachtree Corners Branch, 5570 Spalding Drive, Peachtree Corners, Georgia.

DRAFT REPORT

Compliance Status Report

North Berkeley Lake Road Site (HSI 10844) Fire Station 19 Duluth, Gwinnett County Georgia



Gwinnett County Department of Water Resources

January 9, 2018



Report Findings 391-3-19-.06(3)(b)(10)

The Fire Station 19 property was listed on the Hazardous Site Inventory (HSI) with four additional properties as the North Berkeley Lake Road site (HSI #10844) because of arsenic reported in soil. Fire Station 19 entered Georgia's Voluntary Remediation Program (VRP) and was accepted in the VRP by the Georgia Environmental Protection Division (EPD). Arsenic was delineated in soil and groundwater on the property and the source of the arsenic was presumed to be from agricultural land use in the area based on several lines of evidence.

A study was completed for the site-specific soils to determine the fraction of arsenic in soil that was bioavailable from human uptake in order to establish Risk Reduction Standards (RRSs) for this property. Although the bioavailability test methods had not been approved by the U.S. Environmental Protection Agency (EPA), the methodology was subsequently approved by EPA and accepted by EPD. The RRSs established for this property using the site-specific bioavailability are a Type 4 RRS of 280 mg/kg in soil that is two feet or less in depth and a Type 5 RRS of 990 mg/kg for soil deeper than two feet. The groundwater RRS used is the default Type 1/3 RRS of 10 ug/L.

CDM Smith has concluded that the Fire Station 19 property complies with these RRSs. Work remaining to qualify this property for delisting from the HSI includes: 1) final approval of the Compliance Status Report (CSR); 2) CSR public participation; 3) executing an environmental covenant with EPD; and 4) filing of the environmental covenant with the Gwinnett County Recorder of Deeds.



Certification of Compliance 391-3-19-.06(4)(a)

I certify under penalty of law that this report and all attachments were prepared under my direction in accordance with a system designed to assure that qualified personnel properly gather and evaluate the information submitted. Based on my inquiry of the person or persons who manage the system, or those persons directly responsible for gathering the information, the information submitted is, to the best of my knowledge and belief, true, accurate, and complete. I am aware that there are significant penalties for submitting false information, including the possibility of fine and imprisonment for knowing violations.

Based on my review of the findings of this report with respect to the risk reduction standards of the Rules for Hazardous Site Response, Rule 391- 3-19-.07, I have determined that the Fire Station 19 property is in compliance with Type 1/3 groundwater risk reduction standards and the Type 4 and Type 5 risk reduction standards.

E OL P	January 9, 2018
Signature	Date
	Division Director of Project Controls
Richard Schoeck	Gwinnett County Department of Water Resources
Name	Title

Qualified Groundwater Scientist Certification

I certify that I am a qualified groundwater scientist who has received a baccalaureate or post-graduate degree in the natural sciences or engineering, and have sufficient training and experience in groundwater hydrology and related fields as demonstrated by state registration and completion of accredited university courses that enable me to make sound professional judgments regarding groundwater monitoring and contaminant fate and transport. I further certify that this report was prepared by myself or by a subordinate working under my direction.

11Duffy

January 9, 2018

Signature

Date

James T. Duffey

PG 000899

Name

P.G. Number





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Section 1

Introduction

This Compliance Status Report (CSR) has been prepared by CDM Smith to fulfill the requirements of the Georgia Voluntary Remediation Program (VRP) and the Georgia Hazardous Site Response Rules (391-3-19) for Fire Station 19. The fire station property is included in the North Berkeley Lake Road Site and is listed on Hazardous Site Inventory (HSI) as HSI #10844. This CSR satisfies the reporting requirements of the Rule 391-3-19-.06(3) and includes the required certifications under Rule 391-3-19-.06(4). The CSR submittal deadline is July 15, 2020 (EPD, December 9, 2016).

A site description is provided below followed by a narrative of the compliance activities completed for the property since enrollment in the VRP. **Section 2** of this CSR describes the property assessment activities and the results. The Risk Reduction Standards (RRSs) applicable to this property have been previously accepted by the Georgia Environmental Protection Division (EPD) and are summarized in **Section 3**. CDM Smith's conclusions and recommendations for the property are presented in **Section 4**. In summary, CDM Smith concludes that the Fire Station 19 property complies with the applicable RRSs identified in Section 3. Recommendations for Gwinnett County are listed below.

- 1. Complete the required public notification requirements upon submitting this CSR to EPD.
- 2. Once EPD approves the CSR, develop an environmental covenant (EC) using EPD's Uniform Environmental Covenant (UEC) that places protective restrictions, limitations, and controls on property use.
- 3. Implement protective restrictions/controls identified in the UEC.

Once the CSR has been approved and the recommendations have been fulfilled, EPD will determine whether the Fire Station 19 property is eligible and should be removed from the HSI.

1.1 Property Description

Fire Station 19 is located at 3275 North Berkeley Lake Road North, Duluth, Georgia, in zip code 30096 (**Figure 1-1**). The 3-acre property is zoned M2 – Heavy Industry. The County acquired the property as vacant land in 1989 and the fire station was built in 1997. The current owner is listed as the Gwinnett County Fire Department and the Parcel Identification Number is R6267-028.

As shown on Figure 1-1, four properties in addition to Fire Station 19 have been listed as HSI sites under #10844. All five properties on HSI #10844 were listed by EPD because of arsenic exceeding the reportable quantity. Additional information on these properties and the HSI listings are provided in the VRP narrative in the next section.



The Fire Station property is bounded by North Berkeley Lake Road to the east and the Gwinnet Regional Distribution Center (GRDC) is across North Berkeley Lake Road. Diamond Crystal Brands is to the south, Berkeley Lake Village is to the north, and a storm water retention pond that is part of the Berkeley Lake Village property is to the west. Zoning surrounding the fire station is primarily M2 – Heavy Industry with Berkeley Lake Village being the exception and zoned as OI – Office Institutional District. Residential zoning is to the west across Peachtree Industrial Boulevard and the nearest residence is approximately 1,300 feet from the property.

Access to the fire station is from North Berkeley Lake Road. The property consists of one main building with garage for the fire station, a fueling depot for County vehicles, a pavilion area, driveways, and parking lots (**Figure 1-2**).

1.2 VRP Narrative

A sequential narrative of the events leading up to Fire Station 19 enrolling in the VRP and the actions taken since enrollment are provided below along with the major milestones associated with the surrounding properties, as shown on **Figure 1-3**.

September 2006: The GRDC was listed on the HSI because of arsenic discovered in soil and groundwater and HSI #10844 was first established.

March 2013: Gwinnett County discovered arsenic in soil at the fire station during the planning phase for a sewer line extension project. Soil sampling was completed by the County because of concerns with the GRDC environmental conditions and the potential impacts on the construction project.

May 2013: Gwinnett County submitted a release notification for Fire Station 19 to EPD reporting arsenic in soil (Gwinnett County, May 3, 2013).

August 2013: EPD collected soil samples from several locations in the site vicinity and discovered arsenic in soil exceeding the default RRSs.

December 2013: EPD listed Fire Station 19 on the HSI as part of the North Berkeley Lake Road Site #10844 (EPD, December 6, 2013).

February 2014: EPD lists Diamond Crystal Brands, Suzanna's Kitchen, and Berkeley Lake Village on the HSI (#10844).

October 2014: Fire Station 19 submitted a VRP application and Corrective Action Plan (CAP) to EPD (October 3, 2014).

January 2015: EPD accepted Fire Station 19 into the VRP (EPD, January 15, 2015) and Gwinnett County began to implement the CAP (CDM Smith, October 6, 2014).

March 2015: Gwinnett County submitted the Soil Electron Microprobe Analyses (EMPA) and In Vitro Bioaccessibility (IVBA) reports (CDM Smith(a,b), March 17, 2015). The EMPA testing concluded that the arsenic source was likely from agricultural land use. The IVBA testing



developed alternate RRSs for arsenic in soil based on the property-specific arsenic bioavailability.

May 2015: EPD notified Gwinnett County that the proposed alternate RRSs based on arsenic bioavailability were not acceptable because the methodology had not been approved by the U.S. Environmental Protection Agency (EPA) Headquarters (EPD, May 1, 2015). Gwinnett County subsequently informed EPD that the methodology was under review by EPA Headquarters and would track the review progress (Gwinnett County, June 9, 2015).

April 2017: EPA Headquarters published a validation assessment for arsenic IVBA (EPA(a), April 20, 2017) and methodology (EPA(b), April 20, 2017), which had been used by CDM Smith for Fire Station 19.

May 2017: EPA Superfund Headquarters issues a release of regulatory acceptance for the April 20, 2017 methodology (EPA, May 5, 2017).

July 2017: Gwinnett County submitted the final soil arsenic RRSs (Gwinnett County, July 7, 2017) based on the Hazardous Site Response Rules, previously received comments from EPD, and the EPA-approved IVBA methodology.

September 2017: EPD notified Gwinnett County that the IVBA methodology was acceptable and requested additional information regarding the IVBA sample location selection be provided in the CSR (EPD, September 19, 2017). Gwinnett County subsequently provided the requested information (Gwinnett County, October 9, 2017) and that information is also included in this CSR.

Section 2 Property Assessment

The site is in the Piedmont Physiographic Province, which is characterized by gently rolling hills with rivers and ravines. The property topography is relatively flat with an earthen embankment forming the north property boundary with Berkeley Lake Village. Most precipitation draining from the property is directed into storm sewers that lead to the storm water retention pond to the west. Drainage in the immediate vicinity of the fire station building drains to North Berkeley Lake Road.

The property is underlain by unconsolidated saprolite that was derived from the in-place weathering of the underlying metamorphic bedrock. The saprolite beneath the property is primarily sandy silt. Bedrock has been encountered on the property at depths ranging from approximately 5 feet to over 40 feet.

Groundwater in the site vicinity occurs in the saprolite where this zone is thick enough and within fractures of the crystalline bedrock. Groundwater flow in the saprolite is controlled primarily by topography with groundwater flow mirroring surface drainage patterns. However, the topography of the subsurface bedrock surface can cause deviations in the presumed groundwater flow directions as valleys and ridges on the rock surface can direct groundwater flow.

2.1 Soil Investigation

Fifty-three soil borings were completed across the property in July 2014 using Geoprobe[®] direct-push methods. Prior to sampling, all utilities on the property were located and marked. The boring locations are shown on **Figure 2-1**. A surface soil sample from land surface to a 2-foot depth was collected from all boring locations. Subsurface samples were collected from 50 percent of the locations at the 4-foot depth. Seven duplicate samples were also collected. A total of 85 soil samples were collected including duplicates.

In January 2015, five additional surface soil samples were collected from two areas that exceeded preliminary RRS estimates. The purpose of these samples was to delineate the surface soil exceedance area so that the soil could be excavated along with soil being derived from the sewer line extension project for offsite disposal. These additional locations are shown on Figure 2-1, along with the areas excavated to a two-foot depth.

The soil samples were laboratory analyzed for arsenic by Method 6010C. The full laboratory reports were previously submitted in the CAP (CDM Smith, October 6, 2014). The soil arsenic data are summarized in **Table 2-1**. The subsurface soil arsenic concentrations were statistically higher and more variable than the surface soil arsenic concentrations. Otherwise, the arsenic distributions in soil across the property did not exhibit definitive concentration patterns that would indicate a source or release location.



2.2 Groundwater Investigation

Two groundwater monitor wells were installed in July 2014 at the locations shown on **Figure 2-2**. The wells are 2-inch diameter, schedule 40 polyvinyl chloride and were installed using standard well construction techniques with hollow-stem augers. The well construction details are summarized below.

	MW-1	MW-2
Total Depth	40 feet	35 feet
Screen Interval	35-40 feet	30-35 feet
Sand Interval	33-40 feet	28-35 feet
Bentonite Interval	31-33 feet	26-28 feet
Grout Interval	0-31 feet	0-26 feet
Depth to Water (8/5/14)	21.87 feet	24.91 feet
Sand Interval Bentonite Interval Grout Interval Depth to Water (8/5/14)	33-40 feet 31-33 feet 0-31 feet 21.87 feet	28-35 feet 26-28 feet 0-26 feet 24.91 feet

Groundwater samples were collected from the monitor wells on August 5, 2014, using lowflow purge and sample collection techniques. The samples were analyzed for arsenic by method 6010C. The arsenic results for both samples were below the laboratory practical quantitation limit (PQL) of 0.010 mg/L. The laboratory report for the groundwater results was included in the CAP (CDM Smith, October 6, 2014).

Groundwater levels were measured from the Fire Station 19 wells and from 3 additional wells located on the GRDC property on February 16, 2015. Groundwater surface elevations were calculated from the water level measurements and a potentiometric surface map for saprolite aquifer was prepared, as shown on Figure 2-2. The saprolite potentiometric surface shows a groundwater divide existing in the general vicinity of North Berkley Lake Road with the downgradient direction at Fire Station 19 being to the southwest and to the north-northeast at the GRDC property. These gradient directions are consistent with topography and surface water features, as shown on Figure 1-1.

2.3 In Vitro Bioaccessibility Assays for Arsenic in Soil 2.3.1 IVBA Sample Collection

As proposed in the CAP (CDM Smith, October 6, 2014), soil samples were collected in October 2014 for analyses by IVBA. The sample collection locations are shown on **Figure 2-3**. The location selection criteria used for the IVBA samples are listed below.

- Geographic Coverage One objective was to provide thorough geographic coverage. The sample collection locations shown on Figure 2-3 demonstrate thorough coverage.
- High Arsenic Bias A second objective was to bias results toward higher arsenic concentrations in soil. Summary statistics comparing the delineation results to the arsenic bioavailability results demonstrate a high bias for the IVBA samples. The total arsenic concentrations in the soil used for IVBA testing are summarized in **Table 2-2**. The bioavailability samples are higher in part because they were sieved



prior to analysis, as required by the analytical method, and the IVBA soil arsenic concentrations should not be compared to RRSs for compliance purposes.

 Sample Depths – The bioavailability samples were depth-composite samples collected from land surface to 2 feet below land surface. This depth was selected to best represent soil that is most frequently subject to human exposure.

2.3.2 IVBA Results

The IVBA laboratory analyses were completed in accordance with the method approved by EPA (EPA, May 5, 2017). The analyses were completed by University of Colorado Geological Sciences Laboratory under the direction of Dr. John Drexler, a key contributor to the method development and the EPA method validation. The method release memorandum by EPA and the method are in **Appendix A** and be accessed at <u>https://www.epa.gov/superfund/soil-bioavailability-superfund-sites-guidance</u>.

CDM Smith previously submitted a detailed technical memorandum describing IVBA (CDM Smith(a), March 17, 2015). **Table 2-2** summarizes the IVBA data and relative bioavailability (RBA) results. During the testing, arsenic was extracted from soil using a fluid that had properties that resembled gastrointestinal fluid (i.e., pH 1.5), residence time in the stomach, temperature, and soil to liquid ratio (i.e., relative amounts of soil and gastrointestinal fluid). The amount of arsenic solubilized from the soil into the fluid was measured. The fraction of solubilized arsenic is referred the IVBA. The fraction IVBA was then utilized to calculate the RBA through the empiric correlation model that has been validated by EPA.

Previous calculations by CDM Smith utilized arsenic IVBA and RBA percentages rounded to one decimal point, as appropriate for the methodology issued in May 2017. However, EPA has since issued an errata sheet for the RBA formula that replaces the use of percentages with fractional units that are rounded to two decimal points for IVBA and RBA. The errata are included in Appendix A.

Test results that had no arsenic detection in the extract were not used to calculate the IVBA. If arsenic were present at the detection limit of 0.2 ug/L in the extract, this would produce an IVBA fraction of 0.00004. The IVBA fractions from the extraction solutions with detections ranged from 0.01 to 0.13.

The empiric correlation model used to calculate RBA from IVBA is shown in Table 2-2 and in the method in Appendix A. The updated RBA values calculated according to the EPA errata sheet ranged from 0.04 to 0.13. To be conservative, an RBA value of 13.5 percent was previously selected to support the calculation of RRSs (Gwinnett County, July 7, 2017) prior to the errata. CDM Smith does not propose using the errata-based RBA to increase the RRSs moving forward because the change will not be of any consequence to this CSR and EPD has already approved the RRSs based on the lower RRSs developed from the 13.5% RBA. The RRSs are discussed in greater detail in Section 3.



2.4 Soil Electron Microprobe Analyses 2.4.1 EMPA Sample Collection

EMPA analyses were performed on a subset of ten of the samples collected for IVBA analyses. The EMPA analyses were performed to characterize the arsenic geochemistry in soil at the property to assess potential arsenic sources. A detailed technical memorandum was submitted describing the EMPA testing and results (CDM Smith(b), March 17, 2015). The EMPA sample locations are shown on Figure 2-3. The samples selected for EMPA analyses were the IVBA samples that had the highest arsenic concentrations. In general, EMPA analyses need over 100 mg/kg of arsenic to be useful.

2.4.2 EMPA Results

The arsenic in soil was primarily in the form of iron oxyhydroxides and the iron oxyhydroxides contributed 98 percent of the total arsenic mass in soil. The arsenic concentrations contained in the iron oxyhydroxides were as high as 400 mg/kg. Iron oxyhydroxides have an extremely high capacity to adsorb and retain dissolved arsenic from aqueous solutions. Based on arsenic geochemistry and the EMPA results, CDM Smith concluded that the arsenic in the iron oxyhydroxides was probably derived from either dissolved arsenate or arsenite and subsequent binding in the iron oxyhydroxide solid phase by either of two mechanisms:

- Adsorption of dissolved arsenic onto existing solid iron oxyhydroxide phases in the soil; and/or
- Co-precipitation or adsorption of the arsenic with the precipitation of dissolved aqueous phase iron, forming solid iron oxyhydroxides.

Arsenic in this form is typically not associated with natural mineral occurrences. While many manmade uses of arsenate and arsenite can be identified, uses that could potentially encompass an area as large as the entire Berkeley Lake Road site limits the possible uses. Solid phase arsenic compounds have been used extensively in the past as pesticides and herbicides and include lead arsenate; calcium or sodium arsenate, or arsenite; copper-arsenic compounds; and organic arsenic containing compounds. The solid compounds were typically dissolved in water for direct spraying on fields or applied as a dust.

Calcium arsenate was once a common herbicide and insecticide used for protection of cotton crops. In 1942 alone, approximately 42,000 tons of calcium arsenate were produced in the United States (Kirk-Othmer, 1995). Calcium arsenate was used extensively on cotton in Georgia (Range, W., 1954). The text below was taken from Range and edited to be concise.

...By 1919, the U.S. Department of Agriculture had discovered that dusting cotton with calcium arsenate was an effective Boll weevil control measure. From 1921 to 1923, Boll weevil damage had become disastrous in Georgia, where the annual cotton yield was reduced up to 45 percent. At that time, the State Department of Entomology annually loaded "Peddler Cars" with calcium arsenate, sidetracked a car at virtually every important railroad station, and begged farmers to buy the poison. In 1923, 150 cars of poison were sold in



Georgia. Newspapers also broadcast control measures and bankers appealed to their debtors to accept the offered advice...

Review of historical aerial photographs of the North Berkeley Road site indicates historical agricultural land use. This land use known to have been in place in the earliest available aerial photograph dated 1938 and as recently as 1960.

2.4.3 EMPA Conclusions

CDM Smith has concluded that agricultural use of calcium arsenate is the most likely arsenic source for the North Berkeley Road site. If applied in a solution, some solid phase calcium arsenate probably remained in surface soil due to evaporation of the liquids. Otherwise, calcium arsenate applied as a dust would initially remain in the soil as a solid phase. This solid phase calcium arsenate would then be dissolved by rainfall and migrate deeper into soil. The dissolved arsenic in the infiltrating rainwater was then attenuated with precipitation of iron oxyhydroxides or adsorption onto existing iron oxyhydroxide minerals in the soil. This vadose zone attenuation of arsenic with the iron hydroxides has prevented arsenic from being detected in groundwater at the property.

Section 3 Risk Reduction Standards

The default background concentration for arsenic in soil listed in the Georgia Hazardous Site Response Rules is 20 mg/kg. Both the default Type 1 residential RRS and the default Type 3 non-residential RRS for arsenic in soil are the background concentration of 20 mg/kg. Over 95 percent of the soil samples collected at Fire Station 19 exceeded the background concentration and associated RRSs. As a result, CDM Smith completed a thorough evaluation of potential receptors to guide the development of appropriate and protective RRSs for this property. RRSs have been previously calculated and submitted to EPD for review. Based on EPD comments and our responses, RRSs have been developed that are acceptable to EPD. These RRSs, and summary of the basis for development, are provided below. Detailed discussions of the RRSs and development process can be found in the background documents listed in Section 1.2, VRP Narrative.

3.1 Potential Receptors

A conceptual exposure model is shown on **Figure 3-1** that was developed for the Fire Station 19 property. The model assumes that the arsenic in soil occurs from surface releases, likely associated with agricultural land use. Direct human exposure to arsenic in surface soil is concluded to be a complete exposure pathway for current and future receptors. Outdoor workers and construction workers could potentially be exposed to arsenic in surface soil through dermal contact with soil, ingestion of soil, and inhalation of respirable soil particles on this non-residential property. To be conservative, a trespasser exposure scenario was also included for exposure to arsenic in surface soil.

Based on CDM Smith's current understanding of the arsenic environmental fate and transport mechanisms at the property and soil arsenic data collected for subsurface soil, future construction workers exposure to arsenic in subsurface soil is also possible. Construction work is not currently occurring on the property. The currently available groundwater laboratory data indicate that arsenic is not present in groundwater and the conclusions of the EMPA study provide an explanation as to why arsenic should not be expected in groundwater. However, EPD has requested that the construction worker exposure scenario consider groundwater (EPD, January 16, 2015). Each of the potentially complete exposure scenarios for current and future receptors are to be addressed through the Georgia Hazardous Site Response Rules (391-3-19) RRS development process.



3.2 RRSs

The soil RRSs apply the previously approved arsenic RBA fraction in soil of 0.135, or 13.5 percent. Based on this RBA, the oral slope factor (SF_o) and oral reference dose (RfD_o) for arsenic were adjusted from the default values, as listed below.

•	Default SF _{o-Default} =	1.50E+00 (mg/kg-day) ⁻¹
•	RBA-Adjusted $SF_{o-Adjusted} = SF_{o-Default} \times 0.135 =$	2.03E-01 (mg/kg-day) ⁻¹
•	Default RfD _{o-Default} =	3.00E-04 (mg/kg-day)
•	RBA-Adjusted RfD _{o-Adjusted} = RfD _{o-Default} / $0.135 =$	2.22E-03 (mg/kg-day)

These adjusted values were used to calculate the RRSs for the Fire Station 19 property (**Table 3-1**). The RRS calculations and exposure assumptions used adhere to the Georgia Hazardous Site Response Rules (391-3-19) where appropriate and are also based on comments previously received from EPD during RRS development. The final detailed calculations and assumptions supporting the soil RRS development are in **Appendix B**.

For arsenic in soil from a two-foot depth or less the selected RRS is 280 mg/kg. For arsenic in soil deeper than two feet the selected RRS is 990 mg/kg. For groundwater, the default Type 1/3 RRS of 10 ug/L is selected.

Section 4 Conclusions and Recommendations

CDM Smith concludes that the requirements of the VRP, the applicable Georgia Hazardous Site Response Rules (391-3-19), and the approved CAP for Fire Station 19 have been fulfilled for the Fire Station 19 property.

4.1 Compliance with RRSs

CDM Smith concludes that arsenic in soil and groundwater on the Fire Station 19 property have been sufficiently characterized and delineated; and EPD has concurred with this conclusion. Exceedances of the RRSs discussed in Section 3 have not been observed. CDM Smith concludes that the Fire Station 19 property complies with the RRSs.

4.2 Public Participation

Within seven days of submitting this CSR to EPD, Gwinnett County must fulfill the public participation requirements identified in the Georgia Hazardous Site Response Rules (391-3-19). The requirements of the rules include publishing "*a notice legal organ of the local governments in whose jurisdiction the site is located, announcing that such report is available for inspection by the general public.*"

The public notice must include:

- Site name, address, and owner;
- EPD standard statement requesting public comment prior to a corrective action determination;
- Announcement of a 30-day public comment period with the name, address, and telephone number of the EPD contact person to whom written or oral comments can be made;
- Name, address, and telephone number of the Gwinnett County contact person; and
- The location where the CSR may be viewed and copied.

4.3 Environmental Covenant

Gwinnett County must enter into an EC with the State of Georgia as a consequence of applying the Type 4 and Type 5 RRSs for arsenic in soil at the property. Georgia has developed a UEC that is to be used as model for preparing the EC for the Fire Station 19 property. The EC will identify the location of administrative records, description of contamination and corrective action, and limitations on property use and activities. Gwinnett County anticipates that the following limitations on property use and activities will apply to the Fire Station 19 property.



- Residential, recreational, day care, and/or school uses of the property shall be prohibited;
- The use or extraction of groundwater beneath the property for drinking water or for any other non-remedial purposes shall be prohibited;
- Non-construction activities that are intrusive to the subsurface, such as gardening, shall be prohibited on the property;
- Construction activities that are intrusive to the subsurface that exceed a construction period of one year shall require EPD notification;
- Construction activities that are intrusive to the subsurface that exceed a construction period of one year may require environmental exposure controls, as determined by a qualified professional on a case-by-case basis; and
- Soil removal from the property shall be prohibited, except that soil may be transported to an appropriately-permitted waste disposal facility with prior facility notification of soil conditions and prior acceptance by the facility.

After the EC has been fully executed between EPD and Gwinnett County, the EC must be filed with the Gwinnett County Recorder of Deeds. The Fire Station 19 compliance status is not dependent on engineering controls or a planned environmental monitoring program. As a result, a monitoring and maintenance plan is not required. Gwinnett County will be required to submit an Annual Report certifying the non-residential use of the property and compliance with the use restrictions, limitations, and controls listed above.

4.4 Property Delisting

In consideration of the Fire Station 19 property complying with the RRSs and upon fulfilling the public notice requirements, executing the EC, and recording the EC with the property deed in the records, the Fire Station 19 property will be qualified for delisting from the HSI pursuant to the Georgia Hazardous Site Response Rules (391-3-19).

Section 5

References

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Figures





3,000

Norcross Quadrange, Georgia, 7.5-Minute Topographic Map Series, USGS, 2017. Contour Interval = 20 Feet

0



Scale in Feet 1,000 2,000 Hazardous Site Inventory # 10844 North Berkeley Lake Road Site Properties

Figure 1-1: Site Location







Image Source: Google Earth Image Date: January 2014



North Berkeley Lake Road Site Property Boundaries

0

Scale in Feet 350 700

Figure 1-3: Site Vicinity Map





Image Source: Google Earth Image Date: January 2014



North Berkeley Lake Road Site Property Boundaries

0

Scale in Feet

Figure 2-2 Monitor Well Locations North Berkeley Lake Road Site (HSI 10844)

Fire Station 19 Duluth, Gwinnett County, Georgia



CDM Smith

IVBA - In Vitro Bioaccessibility **EMPA - Electron Microprobe Analysis**



LEGEND

O Incomplete: Current and future scenarios

Complete: Future scenario only

Complete: Current and future scenarios

Figure 3-1: Conceptual Exposure Model



Tables



Arsenic (mg/kg)		
	Surface	Subsurface
Location	Soil	Soil
SB-1/Dup	97	73 / 70
SB-2	75	
SB-3	112	35
SB-4	62	
SB-5	60	
SB-6	47	
SB-7	15	491
SB-8	107	
SB-9	129	105
SB-9-1	130	
SB-9-2	75	
SB-9-3	161	
SB-9-4	96	
SB-10	40	
SB-11	54	33
SB-12	57	
SB-13	37	274
SB-14	153	
SB-14-1	292	
SB-15	98	6
SB-16	62	
SB-17/Dup	31	18 / 29
SB-18	90	
SB-19/Dup	99	53 / 337
SB-20	97	
SB-21	96	10
SB-22	80	
SB-23	81	32
SB-24	83	

	Arsenic	(mg/kg)
Location	Surface Soil	Subsurface Soil
SB-25/Dup	94	85 / 107
SB-26	94	
SB-27	90	31
SB-28	31	
SB-29	72	
SB-30	90	106
SB-31	21	
SB-32	79	61
SB-33	76	
SB-34	96	33
SB-35/Dup	102 / 43	
SB-36	70	103
SB-37	84	
SB-38	98	371
SB-39	98	
SB-40	60	55
SB-41	77	70
SB-42/Dup	67 / 76	
SB-43	80	
SB-44	63	
SB-45	65	45
SB-46	72	
SB-47	65	22
SB-48	63	
SB-49	73	78
SB-50	77	
SB-51	60	218
SB-52/Dup	69 / 88	138
SB-53	51	
Dup - Duplicate sample No sample		

Ar	senic (mg/kg) in So	oil Summary Statistics	
0.5 - 2 Foot Depth		4 Foot Depth	
Mean	73	Mean	106
Median	76	Median	70
Standard Deviation	22	Standard Deviation	119
Minimum	15	Minimum	6
Maximum	112	Maximum	491
Count	53	Count	29
95% UCL of the Mean	79	95% UCL of the Mean	152

Combined Depths

Mean	85
Median	73
Standard Deviation	74
Minimum	6
Maximum	491
Count	82
95% UCL of the Mean	101
LICI Upper confidence lovel	

UCL - Upper confidence level

CDM Smith

Table 2-1: Soil Arsenic Data Summary

		Total Arsenic in Soil (mg/kg)	
	Depth	Delineation	IVBA Sample
Location	(feet)	Results ¹	Results ²
SB-1	0.5 - 2	97.2	99
SB-3	0.5 - 2	112	554
SB-8	0.5 - 2	107	125
SB-9	0.5 - 2	129	99
SB-14	0.5 - 2	153	310
SB-15	0.5 - 2	98.1	34
SB-18	0.5 - 2	89.7	67
SB-19	0.5 - 2	99.3	89
SB-20	0.5 - 2	96.7	89
SB-21	0.5 - 2	96.2	144
SB-24	0.5 - 2	83.3	314
SB-25	0.5 - 2	93.6	207
SB-27	0.5 - 2	90.4	205
SB-30	0.5 - 2	90.1	134
SB-34	0.5 - 2	95.6	103
SB-35	0.5 - 2	102	104
SB-38	0.5 - 2	98.3	62
SB-39	0.5 - 2	97.7	127
SB-43	0.5 - 2	79.9	88
SB-52	0.5 - 2	68.8	195

IVBA Arsonic (ma/ka) in Soil Sun	mary Statistics
TVDA Alsenic (ing/kg/ in Son Sun	iniary statistics
Mean	157
Median	115
Standard Deviation	120
Minimum	34
Maximum	554
Count	20
95% UCL of the Mean	214

IVBA - In Vitro Bioaccessibility

UCL - Upper confidence level

1 - Delineation results are total arsenic in soil

2 - IVBA results are arsenic in mechanically screened soil.

Table 2-2: IVBA Soil Data Summary



Sample	Arsenic in Screened Soil	Soil Extraction Mass	Arsenic in Extract	Fluid Volume	Arsenic IVBA	Arsenic RBA
Code	mg/kg	grams	µg/L	L	fraction	fraction
SB-1	99	1.00152	<0.2	0.1	NA	NA
SB-3	554	1.005	102	0.1	0.02	0.05
SB-8	125	1.0004	<0.2	0.1	NA	NA
SB-9	99	1.0009	<0.2	0.1	NA	NA
SB-14	310	1.00801	<0.2	0.1	NA	NA
SB-15	34	1.00157	<0.2	0.1	NA	NA
SB-18	67	1.00116	<0.2	0.1	NA	NA
SB-19	89	1.00442	<0.2	0.1	NA	NA
SB-20	89	1.00838	<0.2	0.1	NA	NA
SB-21	144	1.00157	<0.2	0.1	NA	NA
SB-24	314	1.0044	39	0.1	0.01	0.04
SB-25	207	1.00789	14	0.1	0.01	0.04
SB-27	205	1.00253	12	0.1	0.01	0.04
SB-30	134	1.00456	<0.2	0.1	NA	NA
SB-34	103	1.00947	139	0.1	0.13	0.13
SB-35	104	1.01199	<0.2	0.1	NA	NA
SB-38	62	1.0053	<0.2	0.1	NA	NA
SB-39	127	1.00568	<0.2	0.1	NA	NA
SB-43	88	1.00497	<0.2	0.1	NA	NA
SB-52	195	1.00547	15	0.1	0.01	0.04

NA - Not Applicable: No calculation for arsenic below the detection limit in the extract. IVBA - In Vitro Bioavailability

RBA - Relative Bioavailability

 IVBA =
 Extract Arsenic x Fluid Volume

 Processed Soil Arsenic x Extraction Soil Mass

RBA = (0.79 x IVBA) + 0.03



Table 2-3: RBA Data Summary

		Applicable	Arsenic	Dementer
Risk Reduction Standards (RRSs)		Depth	Concentration	Remarks
Type 4 Soll	Site-Specific, Non-Residential Outdoor Worker	Surface to 2 feet	280 mg/kg	assumptions
Type 5 Soil ¹ Type 5 Soil ¹	Site-Specific, Construction Worker	2 feet or more Surface to 2 feet	990 mg/kg 3,400 mg/kg	Based on EPD's standard non-residential exposure assumptions with the following EPD-approved exceptions: Noncarcinogen Averaging Time = 1 year Exposure Frequency = 174 days/year Exposure Duration = 1 year Soil Ingestion Rate = 300 mg/day Based on EPD's standard non-residential exposure
				assumptions with the following EPD-approved exceptions: Noncarcinogen Averaging Time = 30 years adult, = 10 years adolescent Body Weight = 45 kg adolescent Exposure Frequency = 100 days/year, 2 hours/day Exposure Duration = 30 years adult, = 10 years adolescent Soil Ingestion Rate = 100 mg/day adult, 200 mg/day adolescent
Type 1/3 Groundwater	Groundwater Default	All	0.01 mg/L	Assumes 100% bioavailability

RRS selected for Fire Station 19 compliance criteria

1 - RRS based on a relative bioavailability fraction of 0.135, or 13.5%, arsenic in soil.

Table 3-1: Risk Reduction Standards


Appendix A IVBA Approval & Methodology





UNITED STATES ENVIRONMENTAL PROTECTION AGENCY WASHINGTON, D.C. 20460

MAY - 5 2017

OFFICE OF SOLID WASTE AND EMERGENCY RESPONSE

NOW THE OFFICE OF LAND AND EMERGENCY MANAGEMENT

MEMORANDUM

SUBJECT: Release of Standard Operating Procedure for an In Vitro Bioaccessibility Assay for Lead and Arsenic in Soil and Validation Assessment of the In Vitro Arsenic Bioaccessibility Assay for Predicting Relative Bioavailability of Arsenic in Soils and Soil-like Materials at Superfund Sites

Schatzi Fitz-James, Acting Director FROM: Assessment and Remediation Division Office of Superfund Remediation and Technology Innovation (OSRTI)

TO: Superfund National Program Managers, Regions 1-10

The purpose of this memorandum is to transmit the Technical Review Workgroup (TRW) for Metals and Asbestos technical documents entitled "Standard Operating Procedure for an In Vitro Bioaccessibility Assay for Lead and Arsenic in Soil" and "Validation Assessment of In Vitro Arsenic Bioaccessibility Assay for Predicting Relative Bioavailability of Arsenic in Soils and Soil-like Materials at Superfund Sites." The Standard Operating Procedure provides an update to EPA Method 1340 (Standard Operating Procedure for an In Vitro Bioaccessibility Assay for Lead in Soil, April 2012, EPA 9200.2-86) by including an assessment of arsenic bioaccessibility. The Validation Assessment Report presents the basis for the Agency's determination that the In Vitro Bioaccessibility Assay (IVBA) method has satisfied the validation and regulatory acceptance criteria for application of the method for arsenic.

EPA Method 1340 was first published as an SW-846 Method by EPA Office of Resource Conservation and Recovery in 2013 for the assessment of lead bioaccessibility as a method to calculate Relative Bioavailability (RBA) and is now regularly used at Superfund sites. Since then, the TRW has worked to incorporate the assessment of arsenic bioaccessibility into this same method. Arsenic and lead are commonly found together at Superfund sites and accurately measuring their RBA has a significant impact on the risk assessment and on the selection of soil cleanup levels. The addition of arsenic to this method allows the arsenic RBA to be measured rapidly and inexpensively. The method does not require the use or sacrifice of animals, and the reduced cost per sample allows risk assessors to obtain a more representative number of soil samples per exposure unit. Additionally, the incorporation of arsenic into the already existing method for lead means that laboratories already have experience performing the assay.

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These two documents can be accessed on the US EPA Superfund Website:

https://www.epa.gov/superfund/soil-bioavailability-superfund-sites-guidance#arsenic. Please contact Matt Lambert at lambert.matthew@epa.gov or 703-603-7174 if you have any questions or concerns.

Attachments:

- "Standard Operating Procedure for an In Vitro Bioaccessibility Assay for Lead and Arsenic in Soil"
- 2. "Validation Assessment of In Vitro Arsenic Bioaccessibility Assay for Predicting Relative Bioavailability of Arsenic in Soils and Soil-like Materials at Superfund Sites."

cc:

James Woolford, OLEM/OSRTI Barbara Hostage, OLEM/OPM Reggie Cheatham, OLEM/OEM Barnes Johnson, OLEM/ORCR David Lloyd, OLEM/OBLR Charlotte Bertrand, OLEM/FFRRO Carolyn Hoskinson, OLEM/OUST Cyndy Mackey, OECA/OSRE Sally Dalzell, OECA/FEO Karen Melvin and Jill Lowe, Region 3 – Lead Region TRW Committee Members NARPM Co-Chairs OHHRRAF Members

Errata Sheet for the

Standard Operating Procedure for an In Vitro Biaccessibility Assay for Lead and Arsenic in Soil

This errata sheet identifies corrections to the Standard Operating Procedure that was published in April 2017.

Location	Previous Text	Correction	Date
Section	The currently preferred models	The currently preferred models	July 6, 2017
12.2,	for predicting RBA from IVBA	for predicting RBA from IVBA	
page 17	for lead (U.S. EPA, 2007b) and	for lead (U.S. EPA, 2007b) and	
	arsenic (Diamond et al., 2016;	arsenic (Diamond et al., 2016;	
	U.S. EPA, 2017) are:	U.S. EPA, 2017) are:	
	$RBA_{lead}(\%) = 0.88 \bullet IVBA(\%) -$	$RBA_{lead}(\%) = (0.88 \bullet IVBA)(\%)$	
	$0.028 \ (R^2 = 0.92)$	-0.028 (R ² = 0.92)	
	$RBA_{\mathrm{arsenic}}(\%) = 0.79 \bullet IVBA(\%)$	$KBA_{\mathrm{arsenic}}(\overset{()}{\overset{()}}{\overset{()}}{\overset{()}{\overset{()}{\overset{()}}{\overset{()}}{\overset{()}}}}}}}}}}$	
	$+3.0 (R^2 = 0.87)$	$1VBA)({}) + 0.03 {3.0} (R^2 =$	
		0.87)	
	where RBA and IVBA are		
	expressed as percentages (not	where RBA and IVBA are	
	fractions).	expressed as fractions	
		percentages (not percentages	
		fractions).	



OLEM 9200.2-164 April 20, 2017 Corrected July 6, 2017

Standard Operating Procedure for an *In Vitro* Bioaccessibility Assay for Lead and Arsenic in Soil

1.0 SCOPE AND APPLICATION

1.1 The purpose of this standard operating procedure (SOP) is to define the proper analytical procedure for the validated in vitro bioaccessibility (IVBA) assay for lead and arsenic in soil (U.S. EPA, 2007b, 2017), to describe the typical working range and limits of the assay, quality assurance (QA), and to indicate potential interferences. The method described herein has been validated only for lead and arsenic in soil, not other contaminants or matrices (e.g., water, air, amended soils, dust, food, etc.) (U.S. EPA, 2007b, 2017).

1.2 The SOP described herein is typically applicable for the characterization of lead and arsenic bioaccessibility in contaminated soil. Users are cautioned that deviations in the method from the assay as described may impact the results and the validity of the method. Users are strongly encouraged to document and report any deviations, as well as any comparisons, with other methods and associated Quality Assurance (QA) requirements.

1.3 This document is intended to be used as a reference for developing site-specific Quality Assurance Project Plans (QAPPs) and Sampling and Analysis Plans (SAPs), but not intended to be used as a substitute for a site-specific QAPP or a detailed SAP or laboratory Standard Operating Procedure. The information contained in this method is provided by EPA as guidance for the analyst and the regulatory community to use in making judgments necessary to generate results that meet the data quality objectives for the intended application.

1.4 Mention of trade names or commercial products does not constitute endorsement or recommended use by U.S. EPA.

1.5 For additional information on method development, see method EPA SW-846-1340 (https://www.epa.gov/sites/production/files/2015-12/documents/1340.pdf) and general information on quality assurance and hazardous waste materials test methods (https://www.epa.gov/hw-sw846/quality-assurance-and-hazardous-waste-test-methods).

2.0 DEFINITIONS

2.1 Bioavailability (BA): The fraction of an ingested dose (i.e., *in vivo*) that crosses the gastrointestinal epithelium and becomes available for distribution to internal target tissues and organs.

2.2 Absolute bioavailability: Bioavailability expressed as a fraction (or percentage) of a dose.

2.3 Relative bioavailability (RBA): The ratio of the bioavailability of a metal in one exposure context (i.e., physical chemical matrix or physical chemical form of the metal) to that in another exposure context. For example, for this method, RBA is defined as the ratio of bioavailability of lead in soil to lead in water.

2.4 Bioaccessibility: An *in vitro* measure of the physiological solubility of the metal that may be available for absorption into the body.

2.5 Batch: A group of analytical and control/QC samples that are extracted simultaneously and is limited to 20 environmental samples in addition to the batch QC samples.

2.6 Phosphate-amended soil: Phosphate rich materials (e.g., fertilizers) applied to lead-contaminated soils.

2.7 Amended soil: *In-situ* remediation approach to sequester a soil contaminant for the purpose of reducing its bioavailability and transport.

2.8 In vitro: Outside the living body and in an artificial environment.

2.9 In vivo: In the living body of an animal.

3.0 BACKGROUND AND METHOD SUMMARY

3.1 Background

Reliable analysis of the potential health hazards from ingestion of lead and arsenic in the environment depends on accurate information on a number of key parameters, including (1) concentration of metal in environmental media (soil, dust, water, food, air, etc.), (2) intake rates of each medium, and (3) the rate and extent of absorption of lead or arsenic (i.e., "bioavailability") from each medium. Knowledge of bioavailability is important because the amount of lead or arsenic that actually enters the blood and body tissues from an ingested medium depends on the physical-chemical properties of both the contaminants and the medium. For example, lead in soil may exist, at least in part, as poorly water-soluble minerals, and may also exist inside particles of inert matrices such as rock or slag of variable size, shape, and association. These chemical and physical properties may tend to influence (usually decrease) the bioavailability of lead when ingested. Thus, equal ingested amounts of different forms of lead in different media may not be of equal health concern.

Since solubilization is usually required for absorption across membranes, poorly soluble forms of metals, with low bioaccessibility, may also have low bioavailability. In certain circumstances, if solubility is the major determinant of absorption at the portal of entry, bioaccessibility may be a predictor of bioavailability. Lead is an example of this, as is further discussed in U.S. EPA (2007b).

In vitro bioaccessibility (IVBA%) =
$$\frac{C_{ext} \cdot V_{ext} \cdot 100}{Soil_{conc} \cdot Soil_{mass}}$$

where:

 C_{ext} = *in vitro* extractable contaminant (i.e., lead or arsenic) in the in vitro extract (mg/L) V_{ext} = extraction solution volume (L) $Soil_{conc}$ = contaminant concentration (i.e., lead or arsenic) in the soil sample being assayed (mg/kg) $Soil_{mass}$ = mass of soil sample being assayed (kg)

The *in vitro* bioaccessibility assay described in this SOP provides a rapid and relatively inexpensive alternative to *in vivo* assays for predicting RBA of lead and arsenic in soils and soil-like materials (i.e., sediments, mining materials). The method, which measures the extent of metal solubilization in an extraction solvent that resembles gastric fluid, is based on the concept that solubilization of metals in gastrointestinal fluid is likely to be an important determinant of bioavailability *in vivo*. The IVBA is used to estimate the *in vivo* RBA. Measurements of IVBA using this assay have been shown to be a reliable predictor of *in vivo* RBA of lead and arsenic in a wide range of soil types and phases from a variety of different sites (U.S. EPA, 2007b, 2017).

3.2 Rationale for Method

Most previous *in vitro* test systems have employed more complex fluid intended to simulate gastric fluid. For example, Medlin (1997) used a fluid that contained pepsin and a mixture of citric, malic, lactic, acetic, and hydrochloric acids. When the bioaccessibility of a series of test substances were compared using 0.4M glycine buffer (pH 1.5) with and without the inclusion of the enzyme and metabolic acids, no significant difference was observed. This indicates that the simplified buffer employed in the procedure is appropriate, even though it lacks some constituents known to be present in gastric fluid.

The dissolution of a contaminant from a test material into the extraction fluid depends on a number of variables including extraction fluid composition, temperature, pH, time, agitation, and solid/fluid ratio. Additional discussion of these procedures is available in U.S. EPA (2007b) and Drexler and Brattin (2007). The following is a discussion of the reasons why the particular variables were established as they were for this IVBA method along with a few caveats:

<u>Temperature</u>. A temperature of 37°C is used because this is approximately the temperature of gastric fluid *in vivo* in humans.

<u>pH.</u> The human gastric pH values tend to range from 1 to 4 during fasting (see U.S. EPA, 2007b, Appendix A). A pH of 1.5 was selected because the highest amounts of lead and arsenic are extracted at pH 1.5, compared with higher pHs (Brattin et al., 2013; U.S. EPA, 2007b).

<u>Extraction Time.</u> The time that ingested material is present in the stomach (i.e., stomachemptying time) is about 1 hour for a child, particularly when a fasted state is assumed (see U.S. EPA, 2007a, Appendix A). Thus, an extraction time of 1 hour should be used. It was found that allowing the bottles to stand at room temperature for up to 4 hours after rotation at 37° C caused no significant variation (<10%) in lead concentration (U.S. EPA, 2007b).

<u>Agitation</u>. If the test material is allowed to accumulate at the bottom of the extraction apparatus, the effective surface area of contact between extraction fluid and the test material may be reduced, which may influence the extent of contaminant solubilization. Depending on which theory of dissolution is relevant (Nernst and Brunner, 1904 or Dankwerts, 1951), agitation will greatly affect either the diffusion layer thickness or the rate of production of fresh surface. Previous workers have noted problems associated with both stirring and argon bubbling methods (Medlin, 1997). Although no systematic comparison of agitation methods was performed, an end-over-end method of agitation is recommended.

<u>Soil/Fluid Ratio and Mass of Test Material.</u> A solid-to-fluid ratio of 1/100 (mass per unit volume) should be used to reduce the effects of metal dissolution when lower ratios (1/5 and 1/25) are used. Tests using NIST Standard Reference Material (SRM) 2710 showed no significant variation (within $\pm 1\%$ of control means) in the fraction of lead extracted with soil masses as low as 0.2 g per 100 mL (U.S. EPA, 2007b). However, use of low masses of test material could introduce variability due to small scale heterogeneity in the sample and/or to weighing errors. Therefore, the final method employs 1.0 g of test material in 100 mL of extraction fluid.

In special cases, the mass of test material may need to be <1.0 g to avoid the potential for saturation of the extraction solution. Tests performed using lead acetate, lead oxide, and lead carbonate indicate that if the bulk concentration of a test material containing these relatively soluble forms of lead exceed approximately 50,000 ppm, the extraction fluid becomes saturated at 37°C and, upon cooling to room temperature and below, lead chloride crystals will precipitate. To prevent precipitation this from occurring, the concentration of lead in the test material should not exceed 50,000 ppm, or the mass of the test material should be reduced to 0.50 ± 0.01 g (U.S. EPA, 2007b). The IVBA extraction has been conducted on soils with arsenic concentrations up to 13,000 ppm (Juhasz et al., 2007). However, studies to determine if the extraction fluid becomes saturated at soil arsenic concentrations >13,000 ppm have not been conducted.

3.3 Summary of Method

After drying and sieving to $150 \,\mu\text{m}$, 1 g of soil sample is rotated with 100 mL (0.1 L) of 0.4 M glycine buffered extraction fluid (pH 1.50) at 37°C for 1 hour. The supernatant is separated from the sample by filtration and analyzed for lead and/or arsenic by an appropriate analytical method (e.g., Method 6010 and Method 6020).

4.0 INTERFERENCES

4.1 Solvents, reagents, glassware, and other sample processing hardware may yield artifacts and/or interferences during sample analysis. All of these materials must be demonstrated to be free from interferences under the conditions of the analysis by analyzing method blanks. Specific selection of reagents may be necessary.

4.2 While the predictive relationship between IVBA and RBA for lead and arsenic has been shown to be applicable to the variety of soil types, anthropogenic sources (e.g., mining operations, orchards), and elemental forms (U.S. EPA, 2007b, 2017), the bioavailability of contaminated soil is influenced by a variety of site-specific considerations and there are limitations when applying both the *in vivo* and *in vitro* assays (U.S. EPA, 2007b). As such, it is essential to identify IVBA samples containing unusual and/or untested forms of either lead or arsenic as potential sources of uncertainty. These samples will help to inform future research to better understand limits on applicability of the methods outlined in this SOP.

4.3 Excess phosphate in the sample medium may result in interference for the measurement of lead. IVBA results for phosphate-treated soils have not been shown to correlate with extraction results from juvenile swine *in vivo* assays (Scheckel et al., 2013). As a result, the methodology discussed in this SOP is not suited for lead in phosphate-amended soils. The role of phosphate on arsenic IVBA and RBA is not clear; however, phosphate amendments should be avoided in arsenic contaminated soils to avoid unintended transport. The impact of other soil amendments (i.e., iron-based or organic [compost] amendments) have not been fully examined to determine if they influence IVBA results relative to *in vivo* data.

4.4 It is not recommended to analyze lead IVBA for soils exceeding a total lead concentration of 50,000 ppm in order to avoid saturation of the extraction fluid, and because risk management decisions are not likely to be improved by analyzing IVBA for soil with concentrations of lead above this level.

4.5 The IVBA extraction for arsenic has been conducted on soils with arsenic concentrations up to 13,000 ppm (Juhasz et al., 2007). However, users should be cautioned that studies to determine if the extraction fluid becomes saturated at soil arsenic concentrations >13,000 ppm have not been conducted.

4.6 Additional information on interferences and potential problems are discussed further in Section 11.

5.0 SAFETY

This method does not address all safety issues associated with its use. The laboratory is responsible for maintaining a safe work environment and a current awareness file of Occupational Safety and Health Administration (OSHA) regulations regarding the safe handling of the

chemicals specified in this method. A reference file of material safety data sheets (MSDSs) should be available to all personnel involved in these analyses.

6.0 EQUIPMENT AND SUPPLIES

This method recommends the use of a water bath (Section 6.1) or an incubated air chamber (Section 6.2).

A statistical comparison (t-test) was made between the NIST SRM data for lead derived from IVBA extractions that were performed by laboratories employing air (incubator type) as the temperature controlling $(37 \pm 2^{\circ}C)$ medium, versus water (aquarium type water bath). The comparison showed that, for this set of results, there was no statistical difference between the two (2) techniques of controlling the temperature of sample bottles during the extraction.

Additional testing to confirm these results was conducted by EPA's NERL and included four *in vitro* scenarios using NIST SRM 2710a (n = 27 for each scenario):

- 1. Water bath + preheated gastric solution
- 2. Water bath + room temperature gastric solution
- 3. Air incubator + preheated gastric solution
- 4. Air incubator + room temperature gastric solution

Results of the t-tests indicate that there was no statistically significant difference in observed mean Pb IVBA values for NIST 2710a SRM between scenarios 1 and 2; 1 and 3; and 2 and 3. The mean Pb IVBA value from scenario 4 (air temperature controlled, gastric solution not- preheated) was slightly lower. Therefore, the mean Pb IVBA value for scenario 4 was statistically different from the other three scenarios (Nelson et al., 2013).

6.1 Water Bath

If the water bath option is used, the specific extraction device is an electric motor (the same motor as is used in the toxicity characteristic leaching procedure (TCLP, Method 1311) driven flywheel, which drives a rotating block situated inside a temperature-controlled water bath (See Figure 1). The extraction device must be capable of holding a capped 125-mL wide-mouth high density polyethylene (HDPE) bottle. The water bath should be filled such that the extraction bottles are completely immersed. Temperature in the water bath should be maintained at $37 \pm 2^{\circ}$ C using an immersion circulator heater, and the water bath temperature should be monitored and recorded. The electric motor must be capable of 30 ± 2 rpm.





6.2 Incubated Air Chamber

If the air incubator option is used, the specific extraction device will rotate the extraction bottles within an incubated air chamber. It must be capable of rotating at 30 ± 2 rpm and designed to hold capped 125-mL wide-mouth HDPE bottles (see Figure 2 for an example of an extraction device in an incubated air chamber). The incubator must be capable of maintaining $37 \pm 2^{\circ}$ C. The temperature inside the incubator should be monitored and recorded.

Figure 2. Example of *In Vitro* Bioaccessibility Extraction Apparatus with Air Incubator.



6.3 HDPE bottles, 125 mL in size, equipped with airtight screw-cap seals should be used. Care should be taken to ensure that the bottles do not leak and to minimize contamination during the extraction procedure.

6.4 Automated temperature compensation (ATC) pH electrode – used for measuring the pH of the extraction fluid both prior to and after the experiment

7.0 REAGENTS AND STANDARDS

7.1 Reagent grade chemicals, at a minimum, should be used in all tests. Unless otherwise indicated, all reagents should conform to the specifications of the Committee on Analytical Reagents of the American Chemical Society (ACS), where such specifications are available. Other grades may be used, provided the reagent is of sufficiently high purity to permit its use without lessening the accuracy of the determination.

7.2 All reagents shall be free of lead and arsenic. For lead, the final extraction fluid shall be tested to confirm that lead concentrations are $<^{1}/_{4}$ (<one-fourth) of the project-required detection limit (PRDL) of 100 µg/L (i.e., less than 25 µg/L lead in the unprocessed reagent blank). For arsenic, the final extraction fluid shall be tested to confirm that arsenic

concentrations are $<\frac{1}{4}$ (<one-fourth) of the project-required detection limit (PRDL) of 100 µg/L (i.e., less than 25 µg/L arsenic in the unprocessed reagent blank).

7.3 Reagent water must be interference free. All references to water in this method refer to reagent water, unless otherwise specified.

7.4 Cleanliness of all materials used to prepare and/or store the extraction fluid and buffer is essential. All glassware and equipment used to prepare standards and reagents shall be properly cleaned, acid washed, and triple-rinsed with deionized water prior to use.

7.5 Extraction fluid – 0.4 M glycine (free base, reagent-grade glycine in deionized water), adjusted to a pH of 1.50 ± 0.05 at 37°C using trace metal-grade concentrated hydrochloric acid (HCl).

7.5.1 Prepare 2 liters (L) of extraction fluid in a volumetric flask (Class A) using American Society for Testing and Materials (ASTM) Type II deionized (DI) water. Record within two significant digits the weight of glycine using an analytical balance and measure 1.9 L of deionized water ± 1 mL in a pre-acid washed flask. Add 60.06 \pm 0.05 grams of glycine (free base) to a flask containing 1.9 L of deionized water. Glycine should be weighed using an analytical balance calibrated daily according to the manufacturer's instructions. Solution can be transferred to a wide-mouth HDPE bottle for ease of handling. Place the HDPE bottle containing the extraction fluid in a water bath at 37°C and heat until the extraction fluid reaches 37°C. Standardize the pH meter using an ATC pH electrode at 37°C or pH buffers maintained at 37°C in the water bath. Add trace metal-grade concentrated HCl (12.1 N) until the solution pH reaches 1.50 \pm 0.05. Bring the solution to a final volume of 2 L (0.4 M glycine).

7.5.2 If the extraction fluid is prepared in advance of the extraction, the extraction fluid must be heated to 37° C and the pH shall be adjusted to 1.5 using trace metal grade concentrated HCl prior to conducting the extraction batch.

8.0 SOIL SAMPLE PREPARATION, PRESERVATION, AND STORAGE

8.1 All test soils should be prepared by drying ($<40^{\circ}$ C) and sieving to $<150 \,\mu$ m. The $<150 \,\mu$ m size fraction was used because this particle size is representative of that which adheres to children's hands (U.S. EPA, 2016). Stainless steel sieves are recommended. Samples should be thoroughly mixed prior to use to ensure homogenization. Mixing and aliquoting of samples using a riffle splitter is recommended. Clean HDPE storage bottles are recommended.

8.2 To perform this assay, soil standards and test soils should be weighed using an analytical balance calibrated daily according to the manufacturer's instructions. Soil samples should be weighed to *four significant digits* (i.e., the nearest 0.0001 gram).

8.3 All samples should be archived after analysis and retained for further analysis for a period of six (6) months. No preservatives or special storage conditions are required.

9.0 QUALITY ASSURANCE/QUALITY CONTROL

Each laboratory should maintain a formal QA program. The laboratory should also maintain records to document the quality of the data generated. Development of in-house QC limits for each method is encouraged. Use of instrument-specific QC limits is encouraged, provided such limits will generate data appropriate for use in the intended application. All data sheets and QC data should be maintained for reference or inspection. The information contained in this method is provided by EPA as guidance to be used by the analyst and the regulatory community in making judgments necessary to generate results that meet the DQOs for the intended application.

9.1 Initial demonstration of proficiency (IDP)

Each laboratory must demonstrate initial proficiency by generating data of acceptable precision and bias for target analytes in a clean matrix. It is recommended that the laboratory repeat the demonstration of proficiency whenever new staff members are trained or significant changes in instrumentation and/or procedures are made.

9.2 Quality assurance for the extraction procedure are as follows (summarized in Table 1 for lead and Table 2 for arsenic):

9.2.1 Reagent blank: Unprocessed (not run through the extraction procedure) extraction fluid should be analyzed for each new batch of extraction fluid. The reagent blank is considered within control limits if its result is less than the lower limit of quantitation (LLOQ). The corrective action for a blank hit above LLOQ should include preparing a new batch of extraction fluid and reprocessing any samples that were prepared with the failing reagent fluid. The reagent blank should be run at a frequency of 1 in 20 samples (minimum of one per batch).

9.2.2 Method blank: Extraction fluid only (i.e., no test soil) is carried through all steps of the method at a frequency of 1 in 20 samples (minimum of 1 per batch). The method blank is considered within control limits if its result is less than the LLOQ. The corrective action for a recovery above the LLOQ should include making a new extraction fluid and reprocessing any samples that were prepared with the failing method blank.

9.2.3 Laboratory Control Sample (LCS): A LCS consisting of a spiked blank should be run once per batch (minimum 1 in 20 samples). The LCS may be spiked with the same source as the calibration standards and needs to be carried through all steps of the rotation procedure. The extraction fluid should be spiked at either 10 mg/L lead or 10 mg/L arsenic. The control limits are 85–115% recovery. The corrective action for outliers should include an analyst review that all dilutions and spike concentrations were performed correctly. If no error is found, either re-extract the samples or flag and narrate the defect and possible bias in the data.

9.2.4 Matrix Spike (MS): A MS should be run once per batch (minimum 1 in 20 samples). The MS should be prepared after extraction and filtration of the supernatant. The matrix spike should be prepared at either 10 mg/L lead and/or 10 mg/L arsenic. The control limits are 75–125% recovery. The corrective action for outliers should include an analyst review that all dilutions and spike concentrations were performed correctly. If no error is found, either re-extract the samples or flag and narrate the defect and possible bias in the data.

9.2.5 Duplicate sample: A duplicate sample should be run once per batch (minimum 1 in 20 samples) and carried through all steps of the method. The relative percent difference (RPD) should be less than 20%. The corrective action for outliers should include either re-extraction of the samples or flagging the data.

9.2.6 Control soils for Lead: The National Institute of Standards and Testing (NIST) standard reference materials (SRMs) 2710a or 2711a (Montana Soil) can be used as control soils. The reference material shall be carried through all steps of the method and analyzed at a frequency of 1 in 20 samples (minimum of 1 per batch). The IVBA is calculated using the equation shown in Section 12.1.

9.2.6.1 NIST SRM 2710a: Analysis of the NIST SRM 2710a for lead should yield a mean IVBA result of 67.5%, with an acceptable range of 60.7–74.2%. The IVBA result in terms of mg/kg should be 3,440 mg/kg, with a range of 3,096–3,785 mg/kg (Shaw Environmental, Inc., 2011). For the lead concentration (Pb_{soil}) in the SRM, the median lead concentration presented in the Addendum to the NIST certificate for leachable concentrations determined using Method 3050 (5,100 mg/kg) should be used (NIST, 2009a).

9.2.6.2 NIST SRM 2711a: Analysis of the NIST SRM 2711a for lead should yield a mean IVBA result of 85.7%, with an acceptable IVBA range of 75.2–96.2%. The IVBA result in terms of mg/kg should be 1,114 mg/kg, with a range of 980–1,249 mg/kg (Shaw Environmental, Inc., 2011). For the lead concentration (Pb_{soil}) in the SRM, the median lead concentration presented in the Addendum to the NIST certificate for leachable concentrations determined using Method 3050 (1,300 mg/kg) should be used (NIST, 2009b).

9.2.7 Control soils for Arsenic

<u>Note</u>: NIST SRM 2711a is not an appropriate control soil for the IVBA assay for arsenic due to the low arsenic concentration.

9.2.7.1 NIST SRM 2710a: Analysis of the NIST SRM 2710a for arsenic should yield a mean IVBA result of 41.0%, with an acceptable IVBA range 32.9–49.1% (Appendix B). For the arsenic concentration (As_{soil}) in NIST 2710a, the median lead concentration presented in the Addendum to the

NIST certificate for leachable concentrations determined using Method 3050 (1,400 mg/kg) should be used (NIST, 2009a).

9.3 Lower limit of quantitation check standard

9.3.1 The laboratory should establish the LLOQ as the lowest point of quantitation which, in most cases, is the lowest concentration in the calibration curve. The LLOQ should be verified by the analysis of at least seven (7) replicate samples, which are spiked at the LLOQ and processed through all preparation and analysis steps of the method. The mean recovery and relative standard deviation of these samples provide an initial statement of precision and accuracy at the LLOQ. In most cases, the mean recovery should be $\pm 35\%$ of the true value and the RSD should be $\leq 20\%$. In-house limits may be calculated when sufficient data points exist. The monitoring of recovery data for the LLOQ check standard over time is useful for assessing precision and bias. Refer to a scientifically valid and published method (such as Chapter 9 of *Quality Assurance of Chemical Measurements* [Taylor, 1987] or the Report of the Federal Advisory Committee on Detection and Quantitation Approaches and Uses in Clean Water Act Programs [http://water.epa.gov/scitech/methods/cwa/det/index.cfm]) for calculating precision and bias for LLOQ.

9.3.2 Ongoing LLOQ verification, at a minimum, is carried out on a quarterly basis to validate quantitation capability at low analyte concentration levels. This verification may be accomplished either with clean control material (e.g., reagent water, method blanks, Ottawa sand, diatomaceous earth, etc.) or a representative sample matrix (free of target compounds). Optimally, the LLOQ should be less than or equal to the desired regulatory action levels based on the stated project-specific requirements.

Analysis	Frequency	Control Limits	Corrective Action
Reagent blank	once per batch (minimum 1 in 20 samples)	<25 µg/L lead	Make new extraction fluid and rerun all analyses.
Method blank	once per batch (minimum 1 in 20 samples)	<50 μg/L lead	Make new extraction fluid and rerun all analyses.
LCS (10 mg/L)	once per batch (minimum 1 in 20 samples)	85–115% recovery	Ensure dilutions and spike concentrations are correct. If no error is found, re-extract the samples or flag the data.
Matrix spike (10 mg/L)	once per batch (minimum 1 in 10 samples)	75–125% recovery	Ensure dilutions and spike concentrations are correct. If no error is found, re-extract the samples or flag the data.
Duplicate sample	once per batch (minimum 1 in 10 samples)	±20% RPD	Re-extract the samples or flag the data.
Control soil (NIST SRMs 2710a and 2711a)	once per batch (minimum 1 in 20 samples)	NIST 2710a mean 67.5% (acceptable range: 60.7–74.2%) NIST 2711a mean 85.7% (acceptable range: 75.2–96.2%)	Re-extract the samples or flag the data.

 Table 1. Recommended Control Limits for Quality Control Samples: Lead

RPD, Relative percent difference

Table 2.	Recommended	Control Limits fo	r Ouality	Control Samples:	Arsenic
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Analysis	Frequency	Control Limits	Corrective Action
Reagent blank	once per batch (minimum 1 in 20 samples)	<25 µg/L arsenic	Make new extraction fluid and rerun all analyses.
Method blank	once per batch (minimum 1 in 20 samples)	<50 μg/L arsenic	Make new extraction fluid and rerun all analyses.
LCS (10 mg/L)	once per batch (minimum 1 in 20 samples)	85–115 % recovery	Ensure dilutions and spike concentrations are correct. If no error is found, re-extract the samples or flag the data.
Matrix spike (10 mg/L)	once per batch (minimum 1 in 10 samples)	75–125% recovery	Ensure dilutions and spike concentrations are correct. If no error is found, re-extract the samples or flag the data.
Duplicate sample	once per batch (minimum 1 in 10 samples)	±20% RPD	Re-extract the samples or flag the data.
Control soil (NIST 2710a)	once per batch (minimum 1 in 20 samples)	NIST 2710a mean 41.0% (acceptable range: 32.9–49.1%)	Re-extract the samples or flag the data.

RPD, Relative percent difference

10.0 CALIBRATION AND STANDARDIZATION

10.1 An automated temperature compensation (ATC) pH electrode shall be used for measuring the pH of the extraction fluid prior and post experiment. Each instrument/ electrode system must be calibrated at a minimum of two points that bracket the expected pH (1.5) of the samples and are approximately two pH units or more apart. Repeat adjustments on successive portions of the two buffer solutions until readings are within 0.05 pH units of the buffer solution value as indicated in SW-846 method 9045D for Soil and Waste pH (http://www.epa.gov/osw/hazard/testmethods/sw846/pdfs/9045d.pdf). The pH meter should be calibrated and checked with standard solutions within the calibration range (e.g., pH = 1 and 2) according to the manufacturer's instructions. After calibration, the meter is ready to analyze samples.

10.2 Thermometers capable of measuring $37 \pm 2^{\circ}C$ are needed.

10.3 The analytical balance should be calibrated daily in accordance with the manufacturer's instructions.

10.4 Pipettes should be calibrated in accordance with the manufacturer's instructions and the laboratory QA plan.

11.0 PROCEDURE

11.1 The extraction fluid for this procedure is 0.4 M glycine (free base, reagent grade glycine in deionized water), adjusted to a pH of 1.50 ± 0.05 at $37 \pm 2^{\circ}$ C using trace metal grade concentrated hydrochloric acid (HCl). See Section 7.5 for extraction fluid preparation details.

11.2 Pre-heat the TCLP extractor water bath OR incubator (See Section 6.0) to 37°C. Record the temperature at the beginning and end of each extraction batch (an example of an extraction data recording form is provided in Appendix A).

11.3 Soil samples should be thoroughly mixed immediately prior to removing aliquots for extraction to ensure homogenization (i.e., rotate sample bottles using X, Y, Z motion).

11.4 The extraction procedure is begun by placing 1.00 ± 0.05 g of sieved test material (<150 µm; U.S. EPA, 2016) into a 125-mL wide-mouth HDPE bottle. **Record weight of soil to** *four significant digits* (i.e., the nearest 0.0001 gram). Care should be taken to ensure that static electricity does not cause soil particles to adhere to the lip or outside threads of the bottle; if necessary, an antistatic brush should be used to eliminate static electricity prior to adding the test substrate.

11.5 Measure 100 ± 0.5 mL of the $37 \pm 2^{\circ}$ C buffered extraction fluid (0.4 M glycine, pH 1.5), using a graduated cylinder or automated dispenser, and transfer extraction fluid to the 125-mL wide-mouth HDPE bottle.

11.6 The bottle should be tightly sealed and then shaken or inverted to ensure that there is no leakage and that no soil is caked on the bottom of the bottle.

11.7 Fill the extractor (TCLP extractor OR rotating extractor inside of a pre-heated incubator, see Section 6.0 for details) with 125-mL bottles containing test materials or Quality Control samples (see Section 7.0). Record start time of rotation.

<u>NOTE</u>: Care should be taken to prevent contamination of the samples during rotation (e.g., getting bath water in the threads around the cap and possibly into the sample when the cap is removed). Precautions that laboratories may consider include but are not limited to: the type of bottle that is used, sealing the samples in plastic freezer bags with air expelled before installing in the water bath extractor, and/or sealing the bottles with tape or Parafilm[®].

11.8 Samples are extracted by rotating the samples at 30 ± 2 rpm for 1 hour.

11.9 After 1 hour, the bottles should be removed from the rotator, dried, and placed upright on the bench top to allow the soil to settle to the bottom.

11.10 A 40-mL sample of supernatant fluid is then removed directly from the extraction bottle into a disposable syringe. After withdrawal of the sample into the syringe, a Luer-Lok attachment fitted with a 0.45- μ m cellulose acetate disk filter (25 mm diameter) is

attached, and the 15 mL aliquot of fluid is filtered through the attachment to remove any particulate matter into a pre-acid washed polypropylene centrifuge tube or other appropriate sample vial for analysis.

11.11 Record the time that the extract is filtered (i.e., extraction is stopped). If the total time elapsed for the extraction and filtration process exceeds 90 minutes, the test must be repeated (i.e., Steps 11.1–11.10).

11.12 Measure and record the pH of fluid remaining in the extraction bottle. If the fluid pH is not within ± 0.5 pH units of the starting pH, the test must be discarded and the sample re-analyzed. In some cases (mainly slag soils), the test material can increase the pH of the extraction buffer, and this could influence the results of the bioaccessibility measurement. To guard against this, the pH of the fluid should be measured at the end of the extraction step (just after a sample was withdrawn for filtration and analysis). If the pH is not within 0.5 pH units of the starting pH (1.5), the sample should be re-analyzed. If the second test also results in an increase in pH of >0.5 units, it is reasonable to conclude that the test material is buffering the solution. In these cases, the test should be repeated using manual pH adjustment during the extraction process, stopping the extraction at 5, 10, 15, and 30 minutes and manually adjusting the pH down to pH 1.5 at each interval by drop-wise addition of HCl.

11.13 Store filtered sample(s) in a refrigerator at $4 \pm 2^{\circ}$ C until they are analyzed. This filtered sample of extraction fluid is then analyzed for lead and/or arsenic by ICP-AES or ICP-MS (U.S. EPA Method 6010C or Method 6020A). For lead, the method detection limit (MDL) in extraction fluid should be approximately 20 µg/L for Method 6010 and 0.1–0.3 µg/L for Method 6020 (U.S. EPA, 2012a, b). For arsenic, the MDL in extraction fluid should be approximately 20–40 µg/L for Method 6010 and 1–5 µg/L for Method 6020.

<u>NOTE</u>: In some cases, high dissolved solids (e.g., Fe oxides) in the extracts may cause nebulizer performance issues by inductively coupled plasma-optical emission spectrometry (ICP-OES) or inductively coupled plasma-mass spectrometry (ICP-MS). If this is encountered, dilution of the extracts tenfold is recommended before analysis. Correct for any dilutions in the calculations. Alternately, a high solids nebulizer may be useful. Graphite furnace atomic absorption spectrophotometry (GFAA) should be avoided due to the high levels of HCl in the extracts.

<u>NOTE</u>: In some cases, the amount of lead present in the sample will begin to saturate the extraction fluid, and the extraction response will cease to be linear. If the concentration of lead in the extract exceeds approximately 500 mg/L (depending on the sample matrix and mineralogy), this upper limit may have been reached. It is not recommended to analyze IVBA for soils exceeding a total lead concentration of 50,000 ppm in order to avoid saturation of the extraction fluid, and because risk management decisions are not likely to be improved by analyzing IVBA for soil with concentrations of lead above this level.

11.14. Examples of an extraction record, gastric extraction fluid preparation record, and an example batch format and IVBA calculation are provided in Appendix A (Tables A1–A3).

11.15. Once received by the laboratory, all samples and extracts should be checked-in, verified, and maintained under standard chain-of-custody (e.g., U.S. EPA, 2012c).

12.0 DATA ANALYSIS AND CALCULATIONS

A split of each solid material (sieved to $<150 \mu$ m) that has been subjected to this extraction procedure should be analyzed for total lead and/or total arsenic concentration using analytical procedures taken from the U.S. EPA SW-846 (U.S. EPA, 2012d) or a non-destructive method such as Instrumental Neutron Activation Analysis. If SW-846 methods are used, the solid material should be acid digested according to SW-846 Method 3050B (December 1996 revision) or 3051A (microwave-assisted digestion, February 2007 revision), and the digestate analyzed for lead and/or arsenic concentrations determined by ICP-AES analysis (Method 6010C, February 2007 revision) or ICP-MS (Method 6020A, February 2007 revision). Note that although SW-846 Method 3050B states a hot plate is acceptable as a heating source, a hot plate should not be used; the heating source should be a block digestor.

12.1 *In vitro* bioaccessibility (IVBA) is calculated and expressed on a percentage basis using the following equation:

In vitro bioaccessibility =
$$\frac{C_{ext} \cdot V_{ext} \cdot 100}{Soil_{conc} \cdot Soil_{mass}}$$

where:

 C_{ext} = *in vitro* extractable contaminant (i.e., lead/arsenic) in the in vitro extract (mg/L) V_{ext} = extraction solution volume (L)

 $Soil_{conc}$ = contaminant concentration (i.e., lead/arsenic) in the soil sample being assayed (mg/kg)

Soil_{mass} = mass of soil sample being assayed (kg)

12.2 In order for an *in vitro* bioaccessibility test system to be useful in predicting the *in vivo* RBA of a test material, it is necessary to empirically establish that a strong correlation exists between the *in vivo* and the *in vitro* results across many different samples. The currently preferred models for predicting RBA from IVBA for lead (U.S. EPA, 2007b) and arsenic (Diamond et al., 2016; U.S. EPA, 2017) are:

 $RBA_{lead} = (0.88 \bullet IVBA) - 0.028 (R^2 = 0.92)$ $RBA_{arsenic} = (0.79 \bullet IVBA) + 0.03 (R^2 = 0.87)$

where RBA and IVBA are expressed as fractions (not percentages). It is important to recognize that use of this equation to calculate RBA from a given IVBA measurement will yield the "typical" RBA value expected for a test material with that IVBA, and the true RBA may be somewhat different (either higher or lower).

12.3 If dilutions were performed, apply the appropriate corrections to the sample values.

13.0 METHOD PERFORMANCE

13.1 Method Performance for Lead. NIST SRMs 2710a and 2711a should be used as control soils for lead. The soil standard will be analyzed at a frequency of 1 in 20 samples (minimum 1 per batch). The NIST SRMs 2710a and 2711a are available from the National Institute of Standards and Technology, Standard Reference Materials Program (<u>http://www.nist.gov/srm/</u>). Acceptable performances of soil standards for lead are shown in Table 3. The calculations for percent Pb IVBA is shown in Section 12.1.

Table 3.	Method Performance for Lead	

Soil Standard Mean mg/kg		Acceptable mg/kg Range	Mean IVBA Result (%)	Acceptable IVBA Range (%)	
NIST 2710a	3,440	3,096–3,785	67.5	60.7-74.2	
NIST 2711a	1,114	980–1,249	85.7	75.2–96.2	

13.2 Method Performance for Arsenic. NIST SRM 2710a should be used as a control soil for arsenic. The soil standard will be analyzed at a frequency of 1 in 20 samples (minimum 1 per batch). The NIST SRM 2710a is available from the National Institute of Standards and Technology, Standard Reference Materials Program (<u>http://www.nist.gov/srm/</u>). Acceptable performances of soil standards for arsenic are shown in Table 4. The calculation for percent As IVBA is shown in Section 12.1.

Table 4.	Method	Performance	for	Arsenic
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Soil Standard	Mean mg/kg Result	Acceptable mg/kg Range	Mean IVBA Result	Acceptable IVBA Range (%)
NIST 2710a	1400	1300–1600	41.0	32.9-49.1

14.0 POLLUTION PREVENTION

14.1 Pollution prevention encompasses any technique that reduces or eliminates the quantity or toxicity of waste at the point of generation. Numerous opportunities for pollution prevention exist in laboratory operations. The EPA has established a preferred hierarchy of environmental management techniques that places pollution prevention as the management option of first choice (SW-846). Whenever feasible, laboratory personnel should use pollution prevention techniques to address their waste generation. When wastes cannot be feasibly reduced at the source, the Agency recommends recycling as the next best option.

14.2 For information about pollution prevention that may be applicable to laboratories and research institutions consult *Less is Better: Laboratory Chemical Management for Waste Reduction* available from the American Chemical Society's Department of Government Relations and Science Policy, 1155 16th Street, NW, Washington, DC 20036, <u>http://www.acs.org</u>.

15.0 WASTE MANAGEMENT

The Environmental Protection Agency requires that laboratory waste management practices are consistent with all applicable rules and regulations. The Agency urges laboratories to protect the air, water, and land by minimizing and controlling all releases from hoods and bench operations, complying with the letter and spirit of any sewer discharge permits and regulations, and by complying with all solid and hazardous waste regulations, particularly the hazardous waste identification rules and land disposal restrictions. For further information on waste management, consult The Waste Management Manual for Laboratory Personnel, available from the American Chemical Society's Department of Government Relations and Science Policy, 1155 16th Street, NW, Washington, DC 20036, (202) 872-4477.

16.0 REFERENCES

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APPENDIX A

IVBA Extraction Forms and Calculation

Table A-1. Example Extraction Record

Date: Sample ID: BATCH No:									
Extraction Flu	id ID: G	lycine & H	[Cl, pH 1.5; §	SRM ID:					
Spike solution Lead and/or As (100 mL total v	concentr Spiking Solume) la	ation: 10 n Solution Vo	ng/L Pb or 1 endor, Lot No spikes")	0 mg/L As b. (X mL of sta	ndard add	ed to X m	L extracti	on solutio	ons
	Sai	nple Prep	aration			Extracti	ion		
Sample ID	Bottle No.	Volume (mL)	Sample mass (g)	Agitation Time (min)	Initial pH	Final pH	Start Temp (C)	End Temp (C)	Total Time ^a (min)
Acceptable Range		100 ± 0.5	1.00 ± 0.01	60 ± 5	1.50 ± 0.5	1.50 ± 0.5	37 ± 2	37 ± 2	≤90
Method Blank	1								
LCS	2								
Control Soil	3								
Sample ID	4								
Sample ID	5								
Sample ID	6								
Sample ID	7								
Sample ID	8								
Sample ID	9								
Sample ID	10								
Sample ID	11								
Sample ID	12								
Regent blank	13								
Reagent blank i ªTime between	s not extr start of ag	acted throught	igh the <i>in vitr</i> I filtration	o process.					

Sample Batch No: Date Prepared:										
Component		Fluid Pro	eparation	Actual						
I I I I I I	Lot ID	1L	2L	Quantity	Comments					
Deionized water	ASTM Type II	0.95 L (approximate)	1.90 L (approximate)							
Glycine	Sigma Lot No.	$30.04 \pm 0.05 g$	$60.08 \pm 0.05 g$							
HCl (12.1N; Tr. Metal)	Fisher Optima	(approximate)	(approximate)							
Final Volume		1.0 L (class A)	2.0 L (class A)							
pH at 37°C	—	1.50 ± 0.05	1.50 ± 0.05							

Table A-2.	Gastric	Extraction	Fluid	Preparation
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Table A3. Example Batch Format and IVBA Calculation

Date:		Sample ID:										
Batch N	lo.											
Extracti	on Fluid	ID: Glycine & HC	Cl, pH 1.5									
SRM II):											
Spike so	olution co	ncentration: 10 m	g/L Pb	1 1 1	1 1 . 37	.	1.0	(100 T	1 1	\ 1 1 1	1 (('1	22)
Lead Sp	oiking Sol	ution Vendor, Lot	No. (X mL of st	andard ad	ded to X m	L extractio	on solution	s (100 mL	total volum	e) labele	d as "spikes	s)
Datab	D - 441 -			Soil	Soil	X 7 - I	X 7 - I			0/	A	CD -f
Batch #	Bottle No.	Туре	Sample ID	(grams)	(kg)	(mL)	(L)	(mg/L)	5011 [PD] (mg/kg)	^{%0} IVBA	Avg % IVBA	SD of IVBA%
Insert	1	Method Blank	Method blank	n/a	n/a	100	0.1			n/a		
No.	2	LCS	LCS	n/a	n/a	100	0.1			n/a		
	3	Control soil	SRM 2710a	1.0019	0.00100	100	0.1	34.24	5100	67		
	4	Sample	Sample1 a	1.0016	0.00100	100	0.1	32.24	5100	63		
	5	Sample	Sample1 b	1.0006	0.00100	100	0.1	33.24	5100	65	64.1	1.4
	6	Matrix spike	Sample + spike	0.9985	0.00100	100	0.1					
	7	Sample	Sample2 a	1.0029	0.00100	100	0.1				Avg of	
	8	Sample	Sample2 b	1.0022	0.00100	100	0.1				Dups	SD
	9	Matrix spike	Sample + spike	1.0028	0.00100	100	0.1					
	10	Sample	Sample3 a	1.0004	0.00100	100	0.1				Avg of	
	11	Sample	Sample3 b	1.0029	0.00100	100	0.1				Dups	SD
	12	Matrix spike	Sample + spike	0.9972	0.00100	101	0.1		n/a	n/a		•
	13	Reagent blank	unprocessed sample	n/a	n/a	100	0.1		n/a	n/a		

% IVBA = (Concentration in IVBA extract mg/L) (0.1 L) (Concentration in solid mg/kg) (weight of sample kg) • 100

APPENDIX B

Provisional Reference Values for Arsenic IVBA of NIST 2710A Standard Reference Material

Consensus values for In Vitro Bioaccessibility (IVBA) of arsenic in soil reference materials (RM) are needed to support the Standard Operating Procedures (SOP) for determination of arsenic IVBA in soil. EPA is currently conducting multi-laboratory evaluations of arsenic IVBA for NIST 2710A and USGS Flat Creek RMs and has conducted similar evaluations of lead IVBA for these RMs. Until the arsenic IVBA evaluations are completed, EPA recommends using the provisional reference values for NIST 2710A in Table B-1. Although the provisional reference values are based on data from only two laboratories, the estimated prediction interval ($\pm 20\%$) is in the range observed for lead IVBA reference values (Table B-2). The data on which the arsenic IVBA reference values are based are provided in Tables B-3 (summary) and B-4 (individual replicates).

Table B-1. Recommended Provisional Reference	e Value for Arsenic IVBA% of NIST 2710A
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Laboratory	Reference Material	Laboratory Analysis	Total Soil Arsenic Method	Units	Number of Replicates	Lower 99% Prediction Limit	Mean	Upper 99% Prediction Limit	PI as Percent of Mean
All Labs ^a	NIST2710A	Arsenic IVBA	NIST Certificate ^b	%	131	32.9	41.0	49.1	±19.8

^aData provided by Karen Bradham (EPA PRD NERL) and John Drexler (University of Colorado). ^bNIST certificate median soil arsenic concentration: 1400 mg/kg.

Table B-2. Reference Values for Lead IVBA% of Standard Reference Materials

Laboratory	Reference Material	Laboratory Analysis	Total Soil Lead Method	Units	Number of Replicates	Lower 99% Prediction Limit	Mean	Upper 99% Prediction Limit	PI as Percent of Mean
QATS Round Robin	NIST2710A	Lead IVBA	NIST Certificate	%	35	60.7	67.5	74.2	±10
QATS Round Robin	NIST2711A	Lead IVBA	NIST Certificate	%	35	75.2	85.7	96.2	±12.3
QATS Round Robin	Flat Creek	Lead IVBA	EPA 3051A	%	30, 35 ^a	56.0	71.0	86.0	±21.1

^aBased on n=35 estimates of total Pb (mg/kg) and 30 estimates of IVBA Pb (mg/kg).

Laboratory ^a	Reference Material	Laboratory Analysis	Total Soil Arsenic Method	Units	Number of Replicates	Lower 99% Prediction Limit	Mean	Upper 99% Prediction Limit	PI as Percent of Mean
EPA NERL	NIST2710A	Arsenic IVBA	NIST Certificate	%	117	33.1	40.8	48.4	± 18.8
U Colorado	NIST2710A	Arsenic IVBA	NIST Certificate	%	14	30.7	43.0	55.2	±28.5
All Labs	NIST2710A	Arsenic IVBA	NIST Certificate	%	131	32.9	41.0	49.1	±19.8

Table B-3. Values for Arsenic IVBA% of NIST 2710A Based Data from Individual Laboratories and Combined Data

^aData provided by Karen Bradham (EPA PRD NERL) and John Drexler (University of Colorado).

Replicate	Laboratory ^a	Soil Mass (g)	Extracted As (mg/L)	Total Soil As ^b (mg/kg)	As IVBA (%)
1	EPA NERL	1.00	5.59	1400	39.9
2	EPA NERL	1.00	5.56	1400	39.6
3	EPA NERL	1.00	5.33	1400	38.0
4	EPA NERL	1.00	5.14	1400	36.7
5	EPA NERL	1.00	6.40	1400	45.6
6	EPA NERL	1.00	6.40	1400	45.6
7	EPA NERL	1.00	5.98	1400	42.7
8	EPA NERL	1.00	6.15	1400	43.9
9	EPA NERL	1.00	5.46	1400	38.9
10	EPA NERL	1.00	5.82	1400	41.4
11	EPA NERL	1.00	6.39	1400	45.5
12	EPA NERL	1.00	5.25	1400	37.5
13	EPA NERL	1.00	5.26	1400	37.6
14	EPA NERL	1.00	5.19	1400	37.1
15	EPA NERL	1.00	5.54	1400	39.5
16	EPA NERL	1.00	5.43	1400	38.8
17	EPA NERL	1.00	5.52	1400	39.3
18	EPA NERL	1.00	5.20	1400	37.0
19	EPA NERL	1.00	5.08	1400	36.3
20	EPA NERL	1.00	5.19	1400	37.0
21	EPA NERL	1.00	5.24	1400	37.4
22	EPA NERL	1.00	6.01	1400	42.9
23	EPA NERL	1.00	5.57	1400	39.7
24	EPA NERL	1.00	5.58	1400	39.6
25	EPA NERL	1.00	5.66	1400	40.4
26	EPA NERL	1.00	5.25	1400	37.4
27	EPA NERL	1.00	5.25	1400	37.5
28	EPA NERL	1.00	5.51	1400	39.4
29	EPA NERL	1.00	4.89	1400	35.0
30	EPA NERL	1.00	5.61	1400	40.0
31	EPA NERL	1.00	5.36	1400	38.2
32	EPA NERL	1.01	5.94	1400	42.1
33	EPA NERL	1.00	5.86	1400	41.8
34	EPA NERL	1.00	5.84	1400	41.6
35	EPA NERL	1.00	4.83	1400	34.4
36	EPA NERL	1.00	5.12	1400	36.5

Table B-4. NIST 2710A Arsenic IVBA Replicate Data Used in Calculation of Provisional Reference Values

Replicate	Laboratory ^a	Soil Mass (g)	Extracted As (mg/L)	Total Soil As ^b (mg/kg)	As IVBA (%)
37	EPA NERL	1.00	5.29	1400	37.7
38	EPA NERL	1.00	5.88	1400	41.9
39	EPA NERL	1.00	5.69	1400	40.6
40	EPA NERL	1.00	5.88	1400	41.8
41	EPA NERL	1.00	5.70	1400	40.6
42	EPA NERL	1.00	5.44	1400	38.8
43	EPA NERL	1.00	5.35	1400	38.2
44	EPA NERL	1.00	5.38	1400	38.3
45	EPA NERL	1.00	5.37	1400	38.3
46	EPA NERL	1.00	5.42	1400	38.7
47	EPA NERL	1.00	5.30	1400	37.9
48	EPA NERL	1.00	5.10	1400	36.3
49	EPA NERL	1.00	6.00	1400	42.7
50	EPA NERL	1.00	5.21	1400	37.1
51	EPA NERL	1.00	5.19	1400	37.0
52	EPA NERL	1.00	6.29	1400	44.8
53	EPA NERL	1.00	5.92	1400	42.1
54	EPA NERL	1.00	5.64	1400	40.1
55	EPA NERL	1.00	5.60	1400	39.9
56	EPA NERL	1.00	5.73	1400	40.8
57	EPA NERL	1.00	5.90	1400	42.0
58	EPA NERL	1.00	5.59	1400	39.9
59	EPA NERL	1.00	5.55	1400	39.5
60	EPA NERL	1.00	5.73	1400	40.7
61	EPA NERL	1.00	5.95	1400	42.4
62	EPA NERL	1.00	5.83	1400	41.6
63	EPA NERL	1.00	5.63	1400	40.2
64	EPA NERL	1.00	5.64	1400	40.2
65	EPA NERL	1.00	6.18	1400	44.1
66	EPA NERL	1.00	5.70	1400	40.6
67	EPA NERL	1.00	5.39	1400	38.3
68	EPA NERL	1.00	5.85	1400	41.6
69	EPA NERL	1.00	6.14	1400	43.7
70	EPA NERL	1.00	6.05	1400	43.1
71	EPA NERL	1.00	6.53	1400	46.6
72	EPA NERL	1.00	6.13	1400	43.7

Table B-4. NIST 2710A Arsenic IVBA Replicate Data Used in Calculation of Provisional Reference Values

Replicate	Laboratory ^a	Soil Mass (g)	Extracted As (mg/L)	Total Soil As ^b (mg/kg)	As IVBA (%)
73	EPA NERL	1.00	6.35	1400	45.3
74	EPA NERL	1.00	6.21	1400	44.2
75	EPA NERL	1.00	5.24	1400	37.3
76	EPA NERL	1.00	5.60	1400	40.0
77	EPA NERL	1.00	6.05	1400	43.1
78	EPA NERL	1.00	5.99	1400	42.6
79	EPA NERL	1.00	5.45	1400	38.9
80	EPA NERL	1.00	5.73	1400	40.8
81	EPA NERL	1.00	5.79	1400	41.2
82	EPA NERL	1.00	5.55	1400	39.5
83	EPA NERL	1.01	6.09	1400	43.1
84	EPA NERL	1.00	5.68	1400	40.4
85	EPA NERL	1.00	5.28	1400	37.6
86	EPA NERL	1.00	5.26	1400	37.5
87	EPA NERL	1.00	5.50	1400	39.2
88	EPA NERL	1.01	5.67	1400	40.2
89	EPA NERL	1.00	5.36	1400	38.2
90	EPA NERL	1.01	5.70	1400	40.5
91	EPA NERL	1.00	5.68	1400	40.4
92	EPA NERL	1.01	5.48	1400	38.8
93	EPA NERL	1.01	5.35	1400	37.9
94	EPA NERL	1.00	5.62	1400	40.0
95	EPA NERL	1.00	5.63	1400	40.1
96	EPA NERL	1.01	5.94	1400	42.0
97	EPA NERL	1.00	6.57	1400	46.9
98	EPA NERL	1.00	5.77	1400	41.2
99	EPA NERL	1.00	6.14	1400	43.8
100	EPA NERL	1.00	6.50	1400	46.5
101	EPA NERL	1.01	6.36	1400	44.9
102	EPA NERL	1.01	6.14	1400	43.5
103	EPA NERL	1.01	6.62	1400	46.7
104	EPA NERL	1.01	6.21	1400	44.0
105	EPA NERL	1.01	6.70	1400	47.5
106	EPA NERL	1.00	6.45	1400	46.1
107	EPA NERL	1.00	5.73	1400	40.8
108	EPA NERL	1.01	5.87	1400	41.7

Table B-4. NIST 2710A Arsenic IVBA Replicate Data Used in Calculation of Provisional Reference Values

Replicate	Laboratory ^a	Soil Mass (g)	Extracted As (mg/L)	Total Soil As ^b (mg/kg)	As IVBA (%)
109	EPA NERL	1.01	5.98	1400	42.5
110	EPA NERL	1.00	6.04	1400	43.0
111	EPA NERL	1.00	5.42	1400	38.6
112	EPA NERL	1.00	5.49	1400	39.1
113	EPA NERL	1.01	6.15	1400	43.6
114	EPA NERL	1.01	6.63	1400	46.9
115	EPA NERL	1.01	5.93	1400	42.0
116	EPA NERL	1.01	6.14	1400	43.5
117	EPA NERL	1.00	6.44	1400	45.9
118	U. Colorado	1.00	5.10	1400	36.3
119	U. Colorado	1.02	5.22	1400	36.7
120	U. Colorado	1.01	5.69	1400	40.3
121	U. Colorado	1.01	6.55	1400	46.5
122	U. Colorado	1.00	6.69	1400	47.7
123	U. Colorado	1.00	6.34	1400	45.1
124	U. Colorado	1.00	6.75	1400	48.2
125	U. Colorado	1.00	6.45	1400	46.1
126	U. Colorado	1.00	6.34	1400	45.2
127	U. Colorado	1.01	6.46	1400	45.8
128	U. Colorado	1.02	5.79	1400	40.4
129	U. Colorado	1.01	5.69	1400	40.3
130	U. Colorado	1.00	5.68	1400	40.4
131	U. Colorado	1.01	6.02	1400	42.4

Table B-4. NIST 2710A Arsenic IVBA Replicate Data Used in Calculation of Provisional Reference Values

^aData provided by Karen Bradham *(EPA ORD NERL) and John Drexler, University of Colorado. ^bNIST certificate median soil arsenic concentration.



Validation Assessment of *In Vitro* Arsenic Bioaccessibility Assay for Predicting Relative Bioavailability of Arsenic in Soils and Soil-like Materials at Superfund Sites

1. Introduction

This report summarizes the basis for the Agency's determination that the IVBA method for arsenic has satisfied the validation and regulatory acceptance criteria for application of the method in an appropriate regulatory context. Validation and regulatory acceptance criteria developed by the U.S. Environmental Protection Agency (U.S. EPA, 2007a), as adapted from the Interagency Coordinating Committee on the Validation of Alternative Methods (ICCVAM, 1997), have been applied to an *in vitro* arsenic bioaccessibility (IVBA) assay described in detail by Brattin et al. (2013). The arsenic IVBA method estimates site-specific relative bioavailability (RBA) of arsenic in soils quickly and inexpensively relative to in vivo methods. The arsenic IVBA assay is well suited for regulatory use in arsenic risk assessment for several reasons: (1) the assay does not sacrifice animals; (2) the reduced cost and analysis time from use of the IVBA assay in place of in vivo RBA assays will facilitate greater numbers of soil samples analyzed at each site to improve representativeness; (3) regulatory acceptance of the arsenic IVBA assay would lower bioavailability assessment costs by enabling simultaneous assessments of RBA for both arsenic and lead using the existing Standard Operating Procedure (SOP) for the IVBA extraction protocol, which has been previously validated for assessment of RBA of lead in soil (U.S. EPA 2009, 2012a); and (4) some of the U.S. EPA Regional laboratories and commercial laboratories have analytical and quality control experience with the SOP gained from use of the identical assay for lead.

2. Validation Assessment of the In Vitro Arsenic Bioaccessibility Assay

This section discusses the validation criteria established in the Agency soil bioavailability guidance (U.S. EPA, 2007a). Criteria for method validation and regulatory acceptance were consolidated because many of the criteria overlap.

2.1. Scientific and regulatory rationale for the test method, including a clear statement of its proposed use, should be available.

The scientific and regulatory rationale for the arsenic IVBA method is presented in the following:

U.S. EPA. (2007a) Guidance for Evaluating the Bioavailability of Metals in Soils for Use in Human Health Risk Assessment. OSWER 9285.7-80. May 2007. Available online at <u>https://semspub.epa.gov/work/11/175333.pdf</u>

U.S. EPA. (2012b) Recommendations for Default Value for Relative Bioavailability of Arsenic in Soil. OSWER 9200.1-113. December 2012. Available online at <u>https://semspub.epa.gov/work/11/175338.pdf</u>

Regulatory and scientific rationale: The Guidance for Evaluating the Bioavailability of Metals in Soils for Use in Human Health Risk Assessment (U.S. EPA, 2007a) articulates the regulatory
rationale for determining the bioavailability of metals from soils when assessing human health risks at hazardous waste sites:

Accounting for potential differences in oral bioavailability of metals in different exposure media can be important to site risk assessment (U.S. EPA, 1989). This is true for all chemicals, but is of special importance for ingested metals. This is because metals can exist in a variety of chemical and physical forms, and not all forms of a given metal are absorbed to the same extent. For example, a metal in contaminated soil may be absorbed to a lesser extent than when ingested in drinking water or food. Thus, if the oral RfD or CSF for a metal is based on studies using the metal administered in water or food, risks from ingestion of the metal in soil might be overestimated. Even a relatively small adjustment in oral bioavailability can have significant impacts on estimated risks and cleanup goals. (U.S. EPA, 2007a)

The *Recommendations for Default Value for Relative Bioavailability of Arsenic in Soil* (U.S. EPA, 2012b) document articulates the regulatory rationale for site-specific assessment of arsenic bioavailability in soils:

The current default assumption for assessing risk from arsenic in soil is that the bioavailability of arsenic in soil is the same as the bioavailability of arsenic in water (relative bioavailability [RBA] soil/water = 100%). However, recent bioavailability studies conducted in animal models show that bioavailability of arsenic in soil is typically less than that of highly water soluble forms of arsenic (e.g., sodium arsenate dissolved in water). This suggests that bioavailability of arsenic in soil will typically be less than that of arsenic dissolved in drinking water (i.e., RBA<100%). At sites where this applies, the default assumption of RBA=100% will result in an overestimation of risk. (U.S. EPA, 2012b)

In general, the Agency (U.S. EPA, 2007a) recommends that efforts be made to collect data that support site-specific estimates, rather than relying on the default value recommended in this memorandum which may not accurately represent arsenic RBA at any specific site. Use of the national default in place of site specific estimates may underestimate or overestimate risk. Where development of site-specific RBA estimates is not feasible (e.g., screening-level assessments), the default value of 60% can be used, recognizing that the default value is an estimate that is not likely to be exceeded at most sites and is preferable to the assumption of an RBA equal to 100%. (U.S. EPA, 2012b)

2.2. Relationship of the test method endpoint(s) to the endpoint of interest must be described.

The endpoint of interest for risk assessment is a prediction of the oral RBA of arsenic in soil (ratio of oral bioavailability of arsenic in soil to that of water-soluble arsenic) based on a measurement of IVBA of arsenic in soil (solubility of arsenic in soil at gastric pH). The test soil sample is assayed for IVBA, and the corresponding RBA is predicted from a regression model relating IVBA and RBA. This same approach has been validated by EPA for predicting RBA of lead in soil from IVBA (U.S. EPA, 2009).

The IVBA assay for predicting RBA of arsenic in soil is the same extraction procedure validated for predicting the RBA of lead in soil (U.S. EPA, 2009, 2012a). In brief, the IVBA assay

consists of incubating a 1 g soil sample with end-over-end mixing in 100 mL of 0.4 M glycine buffer (pH 1.5) for 1 hour at 37°C (body temperature).

The regression model for predicting RBA of arsenic in soil from IVBA is based on a metaanalysis of concordant data from studies in mice and swine (Bradham et al., 2011, 2013; Brattin et al., 2013; Juhasz et al., 2009, 2014a). Data were combined into a validation dataset consisting of paired IVBA and RBA measurements made on 83 soils collected from different sites and mineral types, including mining, smelting, and pesticide or herbicide application (see Section 2.3 for mineral types). Paired measurements of IVBA and RBA for each of the 83 soil samples were included in a weighted linear regression model (Equation 1) in which IVBA and RBA were based on their respective variances (1/variance). The estimated slope is 0.79 ± 0.01 (SE) and intercept is 3.0 ± 0.1 (SE). The equation of the model is:

$$RBA(\%) = 0.79 \cdot IVBA(\%) + 3.0$$
 Eq. (1)

This model explains approximately 87% of the variance in RBA (weight-adjusted $R^2 = 0.87$). The 95% prediction limit for a single RBA measurement was ±19% RBA. A detailed description of the derivation of the regression model is provided in Diamond et al. (2016). This regression model could be updated periodically by incorporating more data sets as they become available.

2.3. A detailed protocol for the test method must be available and should include a description of the materials needed, a description of what is measured and how it is measured, acceptable test performance criteria (e.g., positive and negative control responses), a description of how data will be analyzed, a list of the materials for which the test results are applicable, and a description of the known limitations of the test, including a description of the classes of materials that the test can and cannot accurately assess.

Standard Operating Procedure: The arsenic IVBA assay extraction protocol is the same as SOP 92000.2-86 for the IVBA assay for lead in soil (U.S. EPA, 2012a, 2017). EPA has developed an SOP specifically for arsenic that includes the SOP 09000.2-86 extraction protocol along with the corresponding analytical procedures for measuring arsenic in the soil and soil-like materials and extracts. The IVBA method is included under the validated methods tab on the SW-846 website as Method 1340 for lead, which will be updated to include arsenic.

Aside from the standard laboratory glassware, reagents, supplies, and equipment, the materials needed for the IVBA assay include 0.4 M glycine (free base, reagent-grade glycine in deionized water, adjusted to a pH of 1.50 ± 0.05 at 37° C using trace metal-grade concentrated hydrochloric acid), and either a water bath or an incubated air chamber with sample rotator is necessary for the extraction of the samples at 37° C. In addition, reference standards NIST 2710a SRM or Flat Creek SRM need to be purchased for use as the control soils in the QA/QC samples. These materials and equipment do not require a large investment from laboratories interested in performing the IVBA assay.

The IVBA assay is meant to measure the fraction of the amount of ingested arsenic that would be solubilized at the low pH of the stomach. The samples are sieved at 150 μ m to mimic the fraction of soil that is likely to stick to human hands and thereby be ingested (U.S. EPA, 2016). The samples are then extracted in a 0.4 M glycine solution, pH 1.5 at 37°C for 1 hour with rotation to mimic gastric conditions. Following the extraction by IVBA assay, the concentration

of arsenic in the extraction solution is measured by ICP-MS or ICP-AES. The total concentration of arsenic in the sample is measured by SW-846 Method 3051A.

As part of the quality control/quality assurance for the IVBA assay, the method requires that a set of quality control samples be run in a batch of samples. Quality control samples are reagent blank (extraction fluid that is not run through the extraction procedure), method blank (extraction fluid that has been run through the extraction procedure), laboratory control sample (LCS; extraction fluid spiked with arsenic that is run through the extraction procedure), matrix spike (spiked matrix, e.g., soil, that is run through the extraction procedure), duplicate sample, and control soil. Control limits and frequency for each quality control sample for arsenic are shown in Table 1.

Quality Control Samples	Frequency	Control Limits for Arsenic		
Reagent blank	once per batch (minimum 1 in 20 samples)	<25 µg/L arsenic		
Method blank	once per batch (minimum 1 in 20 samples)	<50 µg/L arsenic		
LCS (10 mg/L)	once per batch (minimum 1 in 20 samples)	85–115% recovery		
Matrix spike (10 mg/L)	once per batch (minimum 1 in 10 samples)	75–125% recovery		
Duplicate sample	once per batch (minimum 1 in 10 samples)	±20% RPD		
NIST 2710a ^a	once per batch (minimum 1 in 20 samples)	32.9–49.1%		

Table 1. Recommended Control Limits for Quality Control Samples for Arsenic

RPD = Relative percent difference

^aAppendix A

The % IVBA for a sample is determined from the analytical results by Equation 2.

$$IVBA(\%) = [(As_{ext} \times V_{ext})/(As_{soil} \times Soil_{mass}) \times 100$$
 Eq. (2)

where:

As_{ext} = mass concentration of arsenic in the IVBA extract (mg/L)

V_{ext} = IVBA extract solution volume (L)

- As_{soil} = total arsenic concentration (as determined by SW-846 Method 3051A or equivalent) (mg/kg)
- Soil_{mass} = mass of soil extracted by IVBA (kg)

Equation 1 is applied to the % IVBA results to determine the % RBA (see section 2.2).

Applicable test materials: Application of the IVBA method SOP is expected to yield predictions of RBA for individual soil samples that fall within the prediction interval of the assay (±19 RBA%). The prediction interval was based on results from various sources, including mining, smelting, or pesticide applications. Although arsenic mineralogy has not been evaluated for all soils in the data set, the following arsenic mineral phases were identified: sorbed As^V and As^{III}, arsenic trioxide, arsenopyrite, lollingite, realgar, scorodite, and a variety

of arsenic-metal oxides (Bradham et al., 2011, 2013, 2015; Brattin et al., 2013; Juhasz et al., 2007). It is possible that some soils may fall outside of the established prediction interval as a result of an unusual arsenic mineralogy or soil composition not represented in the validation dataset. Therefore, whenever a sample is suspected of containing an unusual and/or untested source material or arsenic mineralogy, this should be identified as a potential data gap and source of uncertainty in the resulting prediction of RBA. As additional samples with a variety of new and different arsenic forms are tested by both *in vivo* and *in vitro* methods, the range of applicability of the method should be refined and expanded.

Assay limitations: The following uncertainties may apply to applications of the IVBA assay.

- i. Sample arsenic concentration limits: The arsenic concentrations of soils tested in the development of the regression model relating IVBA and RBA and its associated prediction interval for the IVBA assay ranged from 40 to 13,000 ppm. This validation range should be sufficient for most applications of the methodology. Although there is no basis for predicting what errors would necessarily be introduced into the predictions of RBA if sample concentrations outside this range were used in the IVBA assay, use of such samples without validating comparisons with results of an *in vivo* assay will introduce additional uncertainty into estimates of RBA. However, applications of the IVBA assay to such high arsenic concentrations (e.g., >7,000 ppm) are unlikely to change risk management decisions; thus, this limitation is not a serious constraint for the utility of the method to support cleanup decisions. If additional data suggests modification of the limits, then the Agency will issue additional guidance. In addition, the minimum soil concentration in the sample is determined by that which is measurable in the assay using the SOP.
- **ii. Particle size**: Soil samples in the validation dataset were sieved for particles less than 250 μ m. Particle size can be expected to affect dissolution of arsenic embedded in soil particles (Karna et al., 2017). Therefore, additional uncertainty will be associated with RBA estimates from IVBA assays of soil samples having particle sizes excluded from the validation dataset (i.e., >250 μ m) U.S. EPA recommends a sieving size of <150 μ m to represent the particle fraction having the highest likelihood of incidental ingestion (Ruby and Lowney, 2012; U.S. EPA, 2016). Arsenic IVBA in soils sieved to <250 μ m were not different from IVBA measured in soils sieved to <150 μ m (Karna et al., 2017).
- **iii.** Uncertainty in predicted RBA value: The IVBA assay for arsenic measures IVBA for a test soil and converts this to an estimate of RBA using a regression equation estimated from a meta-analysis of 83 samples. The predicted RBA is the most likely (highest probability) estimate corresponding to the IVBA, but the actual RBA (if measured *in vivo*) might be either higher or lower than the predicted value. The 95% prediction limit for the arsenic IVBA-RBA regression model is relatively narrow in the context of its application to risk assessment, ± 19 RBA%. This means that there will be a 95% probability that individual RBA measurements will be ± 19 of the RBA% predicted from IVBA. In general, the most likely estimate of RBA is the most appropriate value for use in risk assessments because there is an equal probability of the true RBA being above or below the predicted value; however, other values from within the RBA prediction interval could also be evaluated as part of an uncertainty analysis.

Predicting RBA in humans: The IVBA assay was developed to predict arsenic RBA in iv. humans, although there are no data in humans to provide a direct validation of RBA predictions in humans. Therefore, the arsenic IVBA assay was evaluated with estimates of RBA made from studies conducted in two different juvenile swine bioassays and a mouse bioassay. The use of animals for establishing arsenic RBA values to be used in regulatory contexts has several precedents: (1) a national default soil arsenic RBA, to be used when site-specific estimates are not available (it is always better to collect and analyze site-specific data than to rely on a default value), was derived based on a large sample of soil RBA measurements made in mice, monkeys, and swine (U.S. EPA, 2012a,c); (2) an IVBA assay was validated for predicting lead RBA based on soil RBA measurements made in a swine assay (U.S. EPA, 2009); and (3) animal bioassays (e.g., mice, monkeys, swine) remain valid for establishing site-specific soil arsenic and lead RBA, but are not recommended because it is better to run IVBA analyses on many samples (e.g., a statistical sample) than to rely on a smaller number of samples analyzed in animal bioassays (U.S. EPA, 2007b, 2010). Significantly greater costs and time to complete will limit the number of animal bioassays.

Although there is no quantitative support for discerning which animal bioassay provides a more accurate prediction of arsenic RBA in humans, RBA estimates obtained from the mouse and swine assays are in close agreement (Bradham et al., 2013; Juhasz et al., 2014b).

2.4. The extent of within-test variability and the reproducibility of the test within and among laboratories must have been demonstrated. The degree to which sample variability affects this test reproducibility should be addressed.

Within-test variability: Precision of the IVBA protocol was assessed with analyses of soils included in the validation dataset, which included contributions from three laboratories. Each laboratory achieved consistent and relatively low coefficients of variation (CV=standard deviation/mean): 2.1, 4.0, and <5% (Brattin et al., 2013; Diamond et al., 2016).

Inter-laboratory reproducibility: An inter-laboratory comparison of the IVBA was conducted with four participating laboratories: ACZ Laboratories Inc.; EPA Region 7 laboratory; EPA Region 8 laboratory; and University of Colorado at Boulder (Brattin et al., 2013). Each laboratory applied the IVBA method to analyses (in triplicate) of 12 test soils. Average within-laboratory variability (coefficient of variation, CV) ranged from 1.3 to 11.0%. The inter-laboratory coefficient ranged from 2.2 to 15% (mean: 5.4%).

Effects of sample variability: The prediction interval for the IVBA assay was derived based on analysis of 83 soil samples from a variety of site types: mining, smelting, or pesticide application. The IVBA range for the soil samples was 0-80% (mean: 27.2 ± 20 SD). The within-laboratory coefficient of variation for IVBA was <0.05 (Diamond et al., 2016).

2.5. The test method performance must have been demonstrated using reference materials or test materials representative of the types of substances to which the test method will be applied, and should include both known positive and known negative agents.

Performance with reference materials: Precision of the IVBA protocol was assessed with replicate arsenic analyses of standard reference materials (SRMs; National Institute of Standards and Technology [NIST] SRM 2710A) conducted by the EPA Office of Research and

Development National Exposure Research Laboratory [ORD NERL]) over several years (Appendix B). The mean relative percent difference ranged from -10.2 to 9.6% (mean: -0.14 \pm 5.3% SD).

Performance with representative materials: The prediction interval for the IVBA assay was derived based on analysis of samples having a variety of arsenic mineral phases from a variety of different types of sites: mining, smelting, and pesticide application.

2.6. Sufficient data should be provided to permit a comparison of the performance of a proposed substitute test with that of the test it is designed to replace.

The IVBA assay is a cost-effective and time-saving alternative to *in vivo* RBA assays that can improve data quality by increasing the number of samples analyzed while reducing costs and turnaround time. For the dataset used to derive the regression model, the model accounted for approximately 87% of the observed variance in RBA. The 95% prediction interval for the model is ± 19 RBA%, based on 83 soil samples from a variety of site types that are expected to be typical applications of the assay for site risk assessment (mining, smelting, and or pesticide application). The standard errors for the RBA estimates for this sample of 83 soils ranged from 0.2 to 20% (median 2%), and the ratios of the SE to the mean RBA (SE/mean) ranged from 0.02 to 0.48 (median 0.09).

2.7. Data supporting the validity of a test method should be obtained and reported in accordance with Good Laboratory Practices (GLPs).

Data supporting validity of the IVBA assay are reported in detail in a published report (Diamond et al., 2016). Data used in the analysis is provided in Appendix C.

2.8. Data supporting the assessment of the validity of the test method must be available for review.

Data supporting the assessment of the validity of the IVBA assay are available online at <u>http://www.tandfonline.com/doi/full/10.1080/15287394.2015.1134038</u>.

2.9. The methodology and results should have been subjected to independent scientific review.

The arsenic IVBA methodology was reviewed by EPA scientists and evaluated in several peerreviewed publications (Bradham et al., 2011, 2013, 2015; Brattin et al., 2013; Juhasz et al., 2009, 2014a,b). The report describing derivation of the prediction regression model was reviewed by the EPA Office of Superfund Remediation and Technology Innovation (OSRTI) Technical Review Workgroup Bioavailability Committee, EPA ORD peer-review for release of publication, and editorial peer-review for publication (Diamond et al., 2016).

2.10. The method should be time and cost effective.

Costs of assessment of a soil sample using the IVBA assay are expected to range from approximately 10-fold to 100-fold less than the costs of a bioassay. Time requirements for the IVBA assay are expected to range from approximately 10-fold to 50-fold less than that required to conduct an *in vivo* bioassay (i.e., days compared to several weeks). Additional cost and time efficiencies are expected for applications at sites where arsenic and lead are chemicals of interest

because the same IVBA extraction protocol can be used to predict arsenic and lead RBA. These efficiencies can be used to analyze a greater number of samples.

2.11. The method should be one that can be harmonized with similar testing requirements of other agencies and international groups.

Other international efforts (e.g., Australia, Canada, European Union, United Kingdom) are pursuing the development of methods for *in vitro* assessment of RBA of arsenic and of other metals and inorganic contaminants in soil. The IVBA assay is directly applicable to these national and international programs. It satisfies the Bioaccessibility Research Canada (BARC) acceptance criteria for use in risk assessment (BARC, 2016; Koch and Reimer, 2012) and the IVBA assay has been used widely to characterize soil arsenic bioaccessibility; recent examples of international use include reports from Africa, Australia, Canada, China, and Great Britain (Dodd et al., 2013; Ettler et al., 2012; Juhasz et al., 2015; Koch and Reimer 2012; Kribek et al., 2014; Li et al., 2015a,b; Meunier et al., 2010; Morales et al., 2015; Silvetti et al., 2014; Wang et al., 2012; Yang et al., 2015). The meta-analysis that forms the basis for the predictive regression model for RBA included contributors from the United States and Australia (Diamond et al., 2016). Various EPA and non-government laboratories provided data to support the validation.

2.12. The method should be suitable for international acceptance.

The IVBA assay is suitable for international acceptance (see section 2.11 for further discussion).

2.13. The method must provide adequate consideration for the reduction, refinement, and replacement of animal use.

The IVBA assay replaces bioassays and will decrease the use of animals for assessing RBA of arsenic in soil.

3. Summary

The IVBA assay for arsenic has been evaluated against validation criteria established by EPA (U.S. EPA, 2007a) for validation of test methods to be used in a regulatory context. All validation criteria have been satisfied. SOPs have been established and tested for intralaboratory precision and inter-laboratory reproducibility. The quantitative relationship between the IVBA assay output and output from *in vivo* animal bioassays, which the IVBA assay is meant to replace, has been reliably established. The description in the method SOP is expected to yield predictions of RBA that fall within acceptable prediction limits for applications in arsenic site risk assessment. The prediction interval is based on assays of samples collected from a variety of arsenic mineral phases from a variety of different sites and, as a result, the method is expected to be widely applicable to soil typically encountered at arsenic waste sites. Based on this assessment, EPA concludes that the IVBA method is valid for predicting RBA of arsenic in soils in support of site-specific risk assessments. The following regression model is recommended for applications to risk assessment (Equation 1):

The Agency strongly encourages use of this methodology when implemented in context with the decision framework described in its soil bioavailability guidance (U.S. EPA, 2007a).

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APPENDIX A

Provisional Reference Values for Arsenic IVBA of NIST 2710A Standard Reference Material

Consensus values for In Vitro Bioaccessibility (IVBA) of arsenic in soil reference materials (RM) are needed to support the Standard Operating Procedures (SOP) for determination of arsenic IVBA in soil. EPA intends to conduct multi-laboratory evaluations of arsenic IVBA for NIST 2710A and USGS Flat Creek RMs. and has conducted similar evaluations of lead IVBA for these RMs. Until the arsenic IVBA evaluations are completed, EPA recommends using the provisional reference values for NIST 2710A in Table A-1. Although, the provisional reference values are based on data from only two laboratories, the estimated prediction interval ($\pm 20\%$) is in the range observed for lead IVBA reference values (Table A-2). The data on which the arsenic IVBA reference values are based are provided in Tables A-3 (summary) and A-4 (individual replicates).

Laboratory	Reference Material	Laboratory Analysis	Total Soil Arsenic Method	Units	Number of Replicates	Lower 99% Prediction Limit	Mean	Upper 99% Prediction Limit	PI as Percent of Mean
All Labs ^a	NIST2710A	Arsenic IVBA	NIST Certificate ^b	%	131	32.9	41.0	49.1	± 19.8

^aData provided by Karen Bradham (EPA PRD NERL) and John Drexler (University of Colorado) ^bNIST certificate median soil arsenic concentration: 1400 mg/kg

Table A-2. Reference	Values for Lead IVBA%	of Standard Reference Materials
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Laboratory	Reference Material	Laboratory Analysis	Total Soil Lead Method	Units	Number of Replicates	Lower 99% Prediction Limit	Mean	Upper 99% Prediction Limit	PI as Percent of Mean
QATS Round Robin	NIST2710A	Lead IVBA	NIST Certificate	%	35	60.7	67.5	74.2	±10
QATS Round Robin	NIST2711A	Lead IVBA	NIST Certificate	%	35	75.2	85.7	96.2	±12.3
QATS Round Robin	Flat Creek	Lead IVBA	EPA 3051A	%	30, 35 ^a	56.0	71.0	86.0	±21.1

^aBased on n=35 estimates of total Pb (mg/kg) and 30 estimates of IVBA Pb (mg/kg)

Laboratory ^a	Reference Material	Laboratory Analysis	Total Soil Arsenic Method	Units	Number of Replicates	Lower 99% Prediction Limit	Mean	Upper 99% Prediction Limit	PI as Percent of Mean
EPA NERL	NIST2710A	Arsenic IVBA	NIST Certificate	%	117	33.1	40.8	48.4	± 18.8
U Colorado	NIST2710A	Arsenic IVBA	NIST Certificate	%	14	30.7	43.0	55.2	± 28.5
All Labs	NIST2710A	Arsenic IVBA	NIST Certificate	%	131	32.9	41.0	49.1	± 19.8

Table A-3. Values for Arsenic IVBA% of NIST 2710A Based Data from Individual Laboratories and Combined Data

^aData provided by Karen Bradham (EPA PRD NERL) and John Drexler (University of Colorado)

Replicate	Laboratory ^a	Soil Mass (g)	Extracted As (mg/L)	Total Soil As ^b (mg/kg)	As IVBA (%)
1	EPA NERL	1.00	5.59	1400	39.9
2	EPA NERL	1.00	5.56	1400	39.6
3	EPA NERL	1.00	5.33	1400	38.0
4	EPA NERL	1.00	5.14	1400	36.7
5	EPA NERL	1.00	6.40	1400	45.6
6	EPA NERL	1.00	6.40	1400	45.6
7	EPA NERL	1.00	5.98	1400	42.7
8	EPA NERL	1.00	6.15	1400	43.9
9	EPA NERL	1.00	5.46	1400	38.9
10	EPA NERL	1.00	5.82	1400	41.4
11	EPA NERL	1.00	6.39	1400	45.5
12	EPA NERL	1.00	5.25	1400	37.5
13	EPA NERL	1.00	5.26	1400	37.6
14	EPA NERL	1.00	5.19	1400	37.1
15	EPA NERL	1.00	5.54	1400	39.5
16	EPA NERL	1.00	5.43	1400	38.8
17	EPA NERL	1.00	5.52	1400	39.3
18	EPA NERL	1.00	5.20	1400	37.0
19	EPA NERL	1.00	5.08	1400	36.3
20	EPA NERL	1.00	5.19	1400	37.0
21	EPA NERL	1.00	5.24	1400	37.4
22	EPA NERL	1.00	6.01	1400	42.9
23	EPA NERL	1.00	5.57	1400	39.7
24	EPA NERL	1.00	5.58	1400	39.6
25	EPA NERL	1.00	5.66	1400	40.4
26	EPA NERL	1.00	5.25	1400	37.4
27	EPA NERL	1.00	5.25	1400	37.5
28	EPA NERL	1.00	5.51	1400	39.4
29	EPA NERL	1.00	4.89	1400	35.0
30	EPA NERL	1.00	5.61	1400	40.0
31	EPA NERL	1.00	5.36	1400	38.2
32	EPA NERL	1.01	5.94	1400	42.1
33	EPA NERL	1.00	5.86	1400	41.8
34	EPA NERL	1.00	5.84	1400	41.6
35	EPA NERL	1.00	4.83	1400	34.4
36	EPA NERL	1.00	5.12	1400	36.5
37	EPA NERL	1.00	5.29	1400	37.7
38	EPA NERL	1.00	5.88	1400	41.9

Table A-4. NIST 2710A Arsenic IVBA Replicate Data Used in Calculation of Provisional Reference Values

Replicate	Laboratory ^a	Soil Mass (g)	Extracted As (mg/L)	Total Soil As ^b (mg/kg)	As IVBA (%)
39	EPA NERL	1.00	5.69	1400	40.6
40	EPA NERL	1.00	5.88	1400	41.8
41	EPA NERL	1.00	5.70	1400	40.6
42	EPA NERL	1.00	5.44	1400	38.8
43	EPA NERL	1.00	5.35	1400	38.2
44	EPA NERL	1.00	5.38	1400	38.3
45	EPA NERL	1.00	5.37	1400	38.3
46	EPA NERL	1.00	5.42	1400	38.7
47	EPA NERL	1.00	5.30	1400	37.9
48	EPA NERL	1.00	5.10	1400	36.3
49	EPA NERL	1.00	6.00	1400	42.7
50	EPA NERL	1.00	5.21	1400	37.1
51	EPA NERL	1.00	5.19	1400	37.0
52	EPA NERL	1.00	6.29	1400	44.8
53	EPA NERL	1.00	5.92	1400	42.1
54	EPA NERL	1.00	5.64	1400	40.1
55	EPA NERL	1.00	5.60	1400	39.9
56	EPA NERL	1.00	5.73	1400	40.8
57	EPA NERL	1.00	5.90	1400	42.0
58	EPA NERL	1.00	5.59	1400	39.9
59	EPA NERL	1.00	5.55	1400	39.5
60	EPA NERL	1.00	5.73	1400	40.7
61	EPA NERL	1.00	5.95	1400	42.4
62	EPA NERL	1.00	5.83	1400	41.6
63	EPA NERL	1.00	5.63	1400	40.2
64	EPA NERL	1.00	5.64	1400	40.2
65	EPA NERL	1.00	6.18	1400	44.1
66	EPA NERL	1.00	5.70	1400	40.6
67	EPA NERL	1.00	5.39	1400	38.3
68	EPA NERL	1.00	5.85	1400	41.6
69	EPA NERL	1.00	6.14	1400	43.7
70	EPA NERL	1.00	6.05	1400	43.1
71	EPA NERL	1.00	6.53	1400	46.6
72	EPA NERL	1.00	6.13	1400	43.7
73	EPA NERL	1.00	6.35	1400	45.3
74	EPA NERL	1.00	6.21	1400	44.2
75	EPA NERL	1.00	5.24	1400	37.3
76	EPA NERL	1.00	5.60	1400	40.0

Table A-4. NIST 2710A Arsenic IVBA Replicate Data Used in Calculation of Provisional Reference Values

Replicate	Laboratory ^a	Soil Mass (g)	Extracted As (mg/L)	Total Soil As ^b (mg/kg)	As IVBA (%)
77	EPA NERL	1.00	6.05	1400	43.1
78	EPA NERL	1.00	5.99	1400	42.6
79	EPA NERL	1.00	5.45	1400	38.9
80	EPA NERL	1.00	5.73	1400	40.8
81	EPA NERL	1.00	5.79	1400	41.2
82	EPA NERL	1.00	5.55	1400	39.5
83	EPA NERL	1.01	6.09	1400	43.1
84	EPA NERL	1.00	5.68	1400	40.4
85	EPA NERL	1.00	5.28	1400	37.6
86	EPA NERL	1.00	5.26	1400	37.5
87	EPA NERL	1.00	5.50	1400	39.2
88	EPA NERL	1.01	5.67	1400	40.2
89	EPA NERL	1.00	5.36	1400	38.2
90	EPA NERL	1.01	5.70	1400	40.5
91	EPA NERL	1.00	5.68	1400	40.4
92	EPA NERL	1.01	5.48	1400	38.8
93	EPA NERL	1.01	5.35	1400	37.9
94	EPA NERL	1.00	5.62	1400	40.0
95	EPA NERL	1.00	5.63	1400	40.1
96	EPA NERL	1.01	5.94	1400	42.0
97	EPA NERL	1.00	6.57	1400	46.9
98	EPA NERL	1.00	5.77	1400	41.2
99	EPA NERL	1.00	6.14	1400	43.8
100	EPA NERL	1.00	6.50	1400	46.5
101	EPA NERL	1.01	6.36	1400	44.9
102	EPA NERL	1.01	6.14	1400	43.5
103	EPA NERL	1.01	6.62	1400	46.7
104	EPA NERL	1.01	6.21	1400	44.0
105	EPA NERL	1.01	6.70	1400	47.5
106	EPA NERL	1.00	6.45	1400	46.1
107	EPA NERL	1.00	5.73	1400	40.8
108	EPA NERL	1.01	5.87	1400	41.7
109	EPA NERL	1.01	5.98	1400	42.5
110	EPA NERL	1.00	6.04	1400	43.0
111	EPA NERL	1.00	5.42	1400	38.6
112	EPA NERL	1.00	5.49	1400	39.1
113	EPA NERL	1.01	6.15	1400	43.6
114	EPA NERL	1.01	6.63	1400	46.9

Table A-4. NIST 2710A Arsenic IVBA Replicate Data Used in Calculation of Provisional Reference Values

Replicate	Laboratory ^a	Soil Mass (g)	Extracted As (mg/L)	Total Soil As ^b (mg/kg)	As IVBA (%)
115	EPA NERL	1.01	5.93	1400	42.0
116	EPA NERL	1.01	6.14	1400	43.5
117	EPA NERL	1.00	6.44	1400	45.9
118	U. Colorado	1.00	5.10	1400	36.3
119	U. Colorado	1.02	5.22	1400	36.7
120	U. Colorado	1.01	5.69	1400	40.3
121	U. Colorado	1.01	6.55	1400	46.5
122	U. Colorado	1.00	6.69	1400	47.7
123	U. Colorado	1.00	6.34	1400	45.1
124	U. Colorado	1.00	6.75	1400	48.2
125	U. Colorado	1.00	6.45	1400	46.1
126	U. Colorado	1.00	6.34	1400	45.2
127	U. Colorado	1.01	6.46	1400	45.8
128	U. Colorado	1.02	5.79	1400	40.4
129	U. Colorado	1.01	5.69	1400	40.3
130	U. Colorado	1.00	5.68	1400	40.4
131	U. Colorado	1.01	6.02	1400	42.4

Table A-4. NIST 2710A Arsenic IVBA Replicate Data Used in Calculation of Provisional Reference Values

^aData provided by Karen Bradham *(EPA ORD NERL) and John Drexler, University of Colorado ^bNIST certificate median soil arsenic concentration

APPENDIX B

Replicate	IVBA (%)	RPD		
1	42.4	3.9		
2	40.0	-1.9		
3	38.5	-5.7		
4	37.2	-9.2		
5	40.9	0.3		
6	37.6	-8.1		
7	39.5	-3.2		
8	43.7	6.9		
9	42.5	4.1		
10	42.8	4.8		
11	40.9	0.3		
12	39.6	-2.9		
13	38.8	-5.0		
14	40.9	0.3		
15	41.6	2.0		
16	39.0	-4.4		
17	42.5	4.1		
18	36.8	-10.2		
19	43.4	6.2		
20	43.3	6.0		
21	42.5	4.1		
22	42.8	4.8		
23	40.9	0.3		
24	39.9	-2.2		
25	39.6	-2.9		
26	44.9	9.6		
27	38.4	-6.0		
Mean	40.8.	-0.14		
SD	2.2	5.32		
Min	36.8	-10.25		
Maximum	44.9	9.63		

Replicate IVBA results for NIST2710A (March 2010 – January 2015) EPA Office of Research and Development National Exposure Research Laboratory

APPENDIX C

Data Used for Meta-analysis of IVBA Assay for Predicting Oral RBA of Arsenic

		Soil As	IVBA	IVBA SD	RBA	RBA SE	
ID	As Source	(ppm)	(%)	(%)	(%)	(%)	RBA Assay
1	Mining/smelting	676	13.0	0.7	38.1	1.6	Swine UEF
2	Mining/smelting	313	32.5	1.6	52.4	2.0	Swine UEF
3	Pesticide (orchard)	290	21.0	1.1	31.0	4.0	Swine UEF
4	Pesticide (orchard)	388	18.6	0.9	40.8	1.8	Swine UEF
5	Pesticide (orchard)	382	19.4	0.4	48.7	4.7	Swine UEF
6	Pesticide (orchard)	364	30.6	1.5	52.8	2.3	Swine UEF
7	Mining/smelting	234	8.8	0.3	17.8	3.2	Swine UEF
8	Mining/smelting	367	6.0	0.3	23.6	2.4	Swine UEF
9	Mining/smelting	181	50.4	2.5	50.7	5.9	Swine UEF
10	Mining	200	78.0	3.9	60.2	2.7	Swine UEF
11	Mining	3957	11.0	0.6	18.6	0.9	Swine UEF
12	Mining/smelting	590	55.1	2.8	44.1	2.3	Swine UEF
13	Mining/smelting	1400	42.2	0.6	41.8	1.4	Swine UEF
14	Mining/smelting	312	41.8	2.1	40.3	3.6	Swine UEF
15	Mining/smelting	983	33.2	1.7	42.2	3.8	Swine UEF
16	Mining/smelting	390	40.3	0.7	36.7	3.3	Swine UEF
17	Mining/smelting	813	22.0	1.1	23.8	2.4	Swine UEF
18	Mining/smelting	368	18.7	0.9	21.2	2.1	Swine UEF
19	Mining/smelting	516	18.6	0.9	23.5	2.6	Swine UEF
20	Herbicide (railway corridor)	267	57.3	2.2	72.2	19.9	Swine AUC
21	Herbicide (railway corridor)	42	42.7	0.8	41.6	6.6	Swine AUC
22	Herbicide (railway corridor)	1114	17.2	0.4	20.0	9.5	Swine AUC
23	Herbicide (railway corridor)	257	10.5	0.1	10.1	2.5	Swine AUC
24	Herbicide (railway corridor)	751	22.2	0.0	22.5	2.2	Swine AUC
25	Herbicide (railway corridor)	91	80.0	0.3	80.5	6.9	Swine AUC
26	Pesticide (dip site)	713	17.8	0.1	29.3	8.7	Swine AUC
27	Pesticide (dip site)	228	55.4	0.6	43.8	5.6	Swine AUC
28	Mining	807	40.0	0.1	41.7	4.4	Swine AUC
29	Mining	577	3.8	0.0	7.0	2.9	Swine AUC
30	Gossan	190	19.0	0.2	16.4	5.2	Swine AUC
31	Gossan	88	14.0	0.2	12.1	4.9	Swine AUC
32	Pesticide	275	5.7	0.2	10.8	0.7	Swine AUC
33	Pesticide	210	7.7	0.4	12.9	1.2	Swine AUC
34	Pesticide	81	41.7	1.1	6.8	1.2	Swine AUC
35	Pesticide	358	6.5	0.1	10.1	3.5	Swine AUC
36	Pesticide	200	13.1	0.3	10.9	3.9	Swine AUC
37	Pesticide	215	7.2	0.2	18.2	3.8	Swine AUC

		Soil As	IVBA	IVBA SD	RBA	RBA SE	
ID	As Source	(ppm)	(%)	(%)	(%)	(%)	RBA Assay
38	Pesticide	981	9.7	0.2	16.4	3.6	Swine AUC
39	Pesticide	1221	15.1	0.6	15.7	1.9	Swine AUC
40	Mining	949	52.9	0.1	45.8	2.6	Swine AUC
41	Mining	1126	36.9	1.1	30.7	4.1	Swine AUC
42	Mining	1695	38.1	1.3	27.5	0.7	Swine AUC
43	Mining	1306	78.4	0.4	70.5	6.8	Swine AUC
44	Mining	2270	43.5	3.4	36.2	1.5	Swine AUC
45	Mining	244	18.1	0.40	15.5	1.3	Mouse UEF
46	Mining	173	6.8	0.80	14.1	1.2	Mouse UEF
47	Mining	6899	17.5	0.60	14.7	1.0	Mouse UEF
48	Mining	280	53.6	0.20	39.9	1.7	Mouse UEF
49	Mining	4495	8.8	0.10	14.5	1.6	Mouse UEF
50	Mining	448	22.8	0.6	17.2	0.5	Mouse UEF
51	Mining	195	25.7	3.4	18.8	2.7	Mouse UEF
52	Mining/smelting	837	18.2	2.70	11.2	0.3	Mouse UEF
53	Mining/smelting	182	32.9	0.20	26.7	1.8	Mouse UEF
54	Mining/smelting	990	73.1	0.60	48.7	2.4	Mouse UEF
55	Mining/smelting	829	74.3	1.30	49.7	2.1	Mouse UEF
56	Mining/smelting	379	53.2	0.50	51.6	2.1	Mouse UEF
57	Pesticide (orchard)	322	18.8	0.30	26.3	1.4	Mouse UEF
58	Pesticide (orchard)	462	16.1	0.40	35.2	2.0	Mouse UEF
59	Pesticide (orchard)	401	18.0	0.20	20.9	2.2	Mouse UEF
60	Pesticide (orchard)	422	27.9	0.80	35.0	1.8	Mouse UEF
61	Pesticide (orchard)	340	35.4	1.90	33.2	2.4	Mouse UEF
62	Pesticide (orchard)	396	48.1	0.80	46.4	1.4	Mouse UEF
63	Pesticide (dip site)	965	9.0	0.40	21.7	1.5	Mouse UEF
64	Pesticide (dip site)	313	36.4	1.30	29.1	1.7	Mouse UEF
65	Herbicide (railway corridor)	246	47.0	2.10	45.1	2.7	Mouse UEF
66	Herbicide (railway corridor)	108	27.0	0.80	23.8	1.9	Mouse UEF
67	Herbicide (railway corridor)	184	11.9	0.20	23.0	1.8	Mouse UEF
68	Herbicide (railway corridor)	981	54.3	2.50	36.3	1.3	Mouse UEF
69	Mining	573	3.5	0.30	6.4	0.3	Mouse UEF
70	Mining	583	21.2	0.20	14.2	0.3	Mouse UEF
71	Gossan	239	12.3	0.70	20.4	1.9	Mouse UEF
72	Mining	197	21.9	0.20	29.0	2.7	Mouse UEF
73	Mining	884	16.9	0.40	23.2	3.3	Mouse UEF
74	Mining	293	12.3	0.30	17.9	0.7	Mouse UEF
75	Mining	223	17.3	0.10	19.8	1.9	Mouse UEF
76	Mining	494	15.5	0.10	18.0	1.8	Mouse UEF

Data Used for Meta-analysis of IVBA Assay for Predicting Oral RBA of Arsenic

		Soil As	IVBA	IVBA SD	RBA	RBA SE	
ID	As Source	(ppm)	(%)	(%)	(%)	(%)	RBA Assay
77	Mining	738	13.4	3.50	11.2	0.9	Mouse UEF
78	Mining	777	0.0	0.00	4.3	0.7	Mouse UEF
79	Mining	943	0.1	0.00	3.0	0.2	Mouse UEF
80	Mining	898	0.1	0.00	1.9	0.2	Mouse UEF
81	Mining	668	0.0	0.00	3.6	0.3	Mouse UEF
82	Mining/smelting (SRM)	601	54.0	4.10	42.9	1.2	Mouse UEF
83	Mining/smelting (SRM)	1513	41.8	1.70	42.1	1.1	Mouse UEF
84	Mining/smelting (SRM)	879	14.5	0.20	14.6	0.8	Mouse UEF

Data Used for Meta-analysis of IVBA Assay for Predicting Oral RBA of Arsenic

As, arsenic; AUC, area under the curve; ID, sample identification number; IVBA, *in vitro* bioaccessibility; RBA, relative bioavailability; SD, standard deviation; SE, standard error; SRM, standard reference material; UEF, urinary excretion fraction

Appendix B RRS Calculations



Type 4 Risk Reduction Standards (RRSs) pose no significant risk on the basis of <u>SITE</u> <u>SPECIFIC EXPOSURE ASSUMPTIONS</u> for <u>NON-RESIDENTIAL PROPERTIES - OUTDOOR WORKER</u>

RAGS Equation 6 Non-Residential Outdoor Worker - Carcinogenic Effects							
C _{car} mg/kg =	<u>TR x BW x AT_{car} x 365 days/year</u> EF x ED x [(SFo x 10 ⁻⁶ kg/mg x IR _{soil}) + (SF _i x IR _{air} x {1/VF + 1/PEF})]						
Parameter	Definition (units)	Default Value	Source				
C _{car} =	Concentration in soil (mg/kg)	Calculated	Not Applicable				
TR _{A/B}	IRIS Carcinogen Class A/B target excess lifetime cancer risk	1.E-05	HSRA Rules				
TR _C	IRIS Carcinogen Class C target excess lifetime cancer risk	1.E-04	HSRA Rules				
SF _i	Inhalation cancer slope factor (mg/kg-dy) ⁻¹ = IUR x 1,000 x BW 70 kg / IR _{air} 20 m ³ /dy	Chemical-Specific	RSLs				
IUR	Inhalation unit risk (ug/m ³) ⁻¹	Chemical-Specific	RSLs				
SFo	Oral cancer slope factor (mg/kg-dy) ⁻¹	Chemical-Specific	RSLs				
AT_{car}	Averaging Time (yr)	70	HSRA Rules				

	RAGS Equation 7							
	Non-Residential Outdoor Worker - Noncarcinoge	enic Effects						
0	<u>THI x BW x AT_{noncar} x 365 days</u>	/year						
C _{noncar} mg/kg =	ED x EF x [{(1/RfD _o) x 10 ⁻⁶ kg/mg x IR _{soil} } + {(1/RfD _i) x IR _{air} x (1/VF + 1/PEF)}]							
Parameter	Definition (units)	Default Value	Source					
C _{noncar} =	Concentration in soil (mg/kg)	Calculated	Not Applicable					
THI	Target hazard index (none)	1	HSRA Rules					
RfD _o	Oral chronic reference dose (mg/kg-dy)	Chemical-Specific	RSLs					
PfD.	Inhalation chronic reference dose (mg/kg-dy)	Chemical Specific	PSI c					
ΝD	= RfC _i x IR _{air} 20 m ³ /dy / BW 70 kg	Chemical-Specific	NOL3					
RfC _i	Inhalation reference concentration (mg/m ³)	Chemical-Specific	RSLs					
AT _{noncar}	Averaging time (yr)	25	HSRA Rules					

Table B-1 Type 4 Outdoor Worker Soil RRS Equations/Assumptions

North Berkeley Lake Road Site (HSI 10844) Fire Station 19 Duluth, Gwinnett County, Georgia



	Standard Assumptions		
BW	Body weight (kg)	70	HSRA Rules
EF	Exposure frequency (dy/yr)	250	HSRA Rules
ED	Exposure duration (yr)	25	HSRA Rules
IR _{air}	Inhalation rate (m ³ /dy)	20	HSRA Rules
IR _{soil}	Soil ingestion rate (mg/dy)	50	HSRA Rules
PEF	Particulate emission factor (m ³ /kg)	4.63E+09	HSRA Rules

	Soil-to-Air Volatilization Factor (VF)							
VF (m ³ /kg) =	$\frac{(\text{LS x V x DH}) \times (\pi \times \alpha \times T)^{1}}{(\pi \times \alpha \times T)^{1}}$							
	A x 2 x D _{ei} x E x K _{as} x 10 ⁻³ kg/g							
Parameter	Definition (units)	Default Value	Source					
LS	Length of side of contaminated area (m)	45	HSRA Rules					
V	Wind speed in mixing zone (m/s)	2.25	HSRA Rules					
А	Area of contamination (cm ²)	2.03E+07	HSRA Rules					
DH	Diffusion height (m)	2	HSRA Rules					
α	$(D_{ei} \times E)/[E + (\rho_s \times (1-E)/K_{as})] (cm^2/s)$	Calculated	HSRA Rules					
Т	Exposure Interval (s)	7.90E+08	HSRA Rules					
ρ _s	Density of soil solids (g/cm ³)	2.65	HSRA Rules					
D _{ei}	Effective diffusivity (cm ² /s)	D _i x E ^{0.33}	HSRA Rules					
D _i	Molecular Diffusivity (cm²/s)	Chemical-specific	RSLs					
E	Total soil porosity	0.35	HSRA Rules					
K _{as}	Soil-air partition coefficient (g soil/cm ³ air)	(H/K _d) x 41	HSRA Rules					
Н	Henry's Law Constant (atm-m ³ /mole)	Chemical-specific	RSLs					
K _d	Soil-water partition coefficient (cm ³ /g)	K _{OC} x OC	HSRA Rules					
K _{oc}	Organic carbon partition coefficient (cm ³ /g)	Chemical-specific	RSLs					
00	Soil Organic Carbon Content (none)	2.0E-02	HSRA Rules					

HSRA Rules: Georgia Hazardous Response Act Rules, 391-3-19, Appendix III, Media Target Concentrations and Standard Exposure Assumptions.

http://rules.sos.state.ga.us/docs/391/3/19/Appendix%20I-IV.pdf

RAGS: Risk Assessment Guidance for Superfund, Volume I - Human Health Evaluation Manual (Part B, Chapter 3, Development of Risk-Based Preliminary Remediation Goals), U.S. Environmental Protection Agency, December 1991. http://www.epa.gov/oswer/riskassessment/ragsb/pdf/chapt3.pdf

> RSLs: U.S. Environmental Protection Agency Regional Screening Levels, November 2017 https://www.epa.gov/risk/regional-screening-levels-rsls-generic-tables-november-2017

> > Table B-1 Type 4 Outdoor Worker Soil RRS Equations/Assumptions



					Carcino	genic Eff	fects			Nonc	arcinoge	nic Effect	S	
		VF	Car.		SFi	IUR	${\rm SF}_{\rm o}$	C _{car}		RfD_{o}	RfC _i	RfD _i	C _{noncar}	Type 4 RRS
Substance	CAS No.	m ³ /mg	Class	TR	(mg/kg-dy) ⁻¹	(ug/m ³) ⁻¹	(mg/kg-dy) ⁻¹	mg/kg	THI	mg/kg-dy	mg/kg-dy	mg/kg-dy	mg/kg	mg/kg
Arsenic, Default	7440382	NA	А	1.E-05	1.51E+01	4.30E-03	1.50E+00	3.82E+01	1	3.00E-04	1.50E-05	4.29E-06	6.13E+02	3.82E+01
Arsenic, RBA Adjusted	7440382	NA	A	1.E-05	1.51E+01	4.30E-03	2.03E-01	2.83E+02	1	2.22E-03	1.50E-05	4.29E-06	4.54E+03	2.83E+02

Data Input Database look up values Spreadsheet calculation

NA - Not Applicable, applies to inhalation for nonvolatile substances and substances not Classified as Class A, B, or C carcinogens.

	SFo	RfD_{o}
	(mg/kg-dy) ⁻¹	mg/kg-dy
Arsenic Default	1.50E+00	3.00E-04
Arsenic, RBA Adjusted	2.03E-01	2.22E-03

SF_o, RBA Adjusted = SF_o Default x RBA RfD_o, RBA Adjusted = RfD_o Default / RBA RBA = 0.0135 or 13.5%

Table B-2 Type 4 Outdoor Worker Soil RRSs

North Berkeley Lake Road Site (HSI 10844) Fire Station 19 Duluth, Gwinnett County, Georgia



Type 5 Risk Reduction Standards (RRSs) pose no significant risk on the basis of <u>SITE</u> <u>SPECIFIC EXPOSURE ASSUMPTIONS</u> for <u>NON-RESIDENTIAL PROPERTIES - CONSTRUCTION WORKER</u>

RAGS Equation 6 Non-Residential Construction Worker - Carcinogenic Effects							
C _{car} mg/kg =	= <u>TR x BW x AT_{car} x 365 days/year</u> EF x ED x [(SFo x 10 ⁻⁶ kg/mg x IR _{soil}) + (SF _i x IR _{air} x {1/VF + 1/PEF})]						
Parameter	Definition (units)	Default Value	Source				
C _{car} =	Concentration in soil (mg/kg)	Calculated	Not Applicable				
TR _{A/B}	IRIS Carcinogen Class A/B target excess lifetime cancer risk	1.E-05	HSRA Rules				
TR_{C}	IRIS Carcinogen Class C target excess lifetime cancer risk	1.E-04	HSRA Rules				
SF _i	Inhalation cancer slope factor (mg/kg-dy) ⁻¹ = IUR x 1,000 x BW 70 kg / IR _{air} 20 m ³ /dy	Chemical-Specific	RSLs				
IUR	Inhalation unit risk (ug/m ³) ⁻¹	Chemical-Specific	RSLs				
SFo	Oral cancer slope factor (mg/kg-dy) ⁻¹	Chemical-Specific	RSLs				
AT _{car}	Averaging Time (yr)	70	HSRA Rules				

RAGS Equation 7 Non-Residential Construction Worker - Noncarcinogenic Effects								
C ma/ka =	THI x BW x AT _{noncar} x 365 days	THI x BW x AT _{noncar} x 365 days/year						
Unoncar mg/kg –	ED x EF x [{(1/RfD _o) x 10 ⁻⁶ kg/mg x IR _{soil} } + {(1/RfD _i) x IR _{air} x (1/VF +	1/PEF)}]					
Parameter	Definition (units)	Default Value	Source					
C _{noncar} =	Concentration in soil (mg/kg)	Calculated	Not Applicable					
THI	Target hazard index (none)	1	HSRA Rules					
RfD _o	Oral chronic reference dose (mg/kg-dy)	Chemical-Specific	RSLs					
RfD.	Inhalation chronic reference dose (mg/kg-dy)		RSIs					
	= RfC _i x IR _{air} 20 m ³ /dy / BW 70 kg	onemical-opecific	NOL3					
RfC _i	Inhalation reference concentration (mg/m ³)	Chemical-Specific	RSLs					
AT _{noncar}	Averaging time (yr)	1	HSRA Rules					

Table B-3 Type 5 Construction Worker Soil RRS Equations/Assumptions

North Berkeley Lake Road Site (HSI 10844) Fire Station 19 Duluth, Gwinnett County, Georgia



	Standard Assumptions		
BW	Body weight (kg)	70	HSRA Rules
EF	Exposure frequency (dy/yr)	174	Site-Specific
ED	Exposure duration (yr)	1	Site-Specific
IR _{air}	Inhalation rate (m ³ /dy)	20	HSRA Rules
IR _{soil}	Soil ingestion rate (mg/dy)	330	Site-Specific
PEF	Particulate emission factor (m ³ /kg)	4.63E+09	HSRA Rules

	Soil-to-Air Volatilization Factor (VF)							
VF (m ³ /kg) =	VF (m ³ /kg) = $\frac{(LS \times V \times DH) \times (\pi \times \alpha \times T)^{1/2}}{(LS \times V \times DH) \times (\pi \times \alpha \times T)^{1/2}}$							
	A x 2 x D _{ei} x E x K _{as} x 10 ^{-°} kg/g							
Parameter	Definition (units)	Default Value	Source					
LS	Length of side of contaminated area (m)	45	HSRA Rules					
V	Wind speed in mixing zone (m/s)	2.25	HSRA Rules					
А	Area of contamination (cm ²)	2.03E+07	HSRA Rules					
DH	Diffusion height (m)	2	HSRA Rules					
α	$(D_{ei} \times E)/[E + (\rho_s \times (1-E)/K_{as})] (cm^2/s)$	Calculated	HSRA Rules					
Т	Exposure Interval (s)	7.90E+08	HSRA Rules					
ρ _s	Density of soil solids (g/cm ³)	2.65	HSRA Rules					
D _{ei}	Effective diffusivity (cm ² /s)	D _i x E ^{0.33}	HSRA Rules					
D _i	Molecular Diffusivity (cm²/s)	Chemical-specific	RSLs					
E	Total soil porosity	0.35	HSRA Rules					
K _{as}	Soil-air partition coefficient (g soil/cm ³ air)	(H/K _d) x 41	HSRA Rules					
Н	Henry's Law Constant (atm-m ³ /mole)	Chemical-specific	RSLs					
K _d	Soil-water partition coefficient (cm ³ /g)	K _{OC} x OC	HSRA Rules					
K _{oc}	Organic carbon partition coefficient (cm ³ /g)	Chemical-specific	RSLs					
00	Soil Organic Carbon Content (none)	2.0E-02	HSRA Rules					

HSRA Rules: Georgia Hazardous Response Act Rules, 391-3-19, Appendix III, Media Target Concentrations and Standard Exposure Assumptions.

http://rules.sos.state.ga.us/docs/391/3/19/Appendix%20I-IV.pdf

RAGS: Risk Assessment Guidance for Superfund, Volume I - Human Health Evaluation Manual (Part B, Chapter 3, Development of Risk-Based Preliminary Remediation Goals), U.S. Environmental Protection Agency, December 1991. <u>http://www.epa.gov/oswer/riskassessment/ragsb/pdf/chapt3.pdf</u>

> RSLs: U.S. Environmental Protection Agency Regional Screening Levels, November 2017 https://www.epa.gov/risk/regional-screening-levels-rsls-generic-tables-november-2017

> > Page 2 of 2

Table B-3 Type 5 Construction Worker Soil RRS Equations/Assumptions North Berkeley Lake Road Site (HSI 10844)

Fire Station 19

Duluth, Gwinnett County, Georgia



					Carcinogenic Effects				Noncarcinogenic Effects					
		VF	Car.		SF _i	IUR	SF。	C _{car}		RfD _o	RfC _i	RfD _i	Cnoncar	Type 4 RRS
Substance	CAS No.	m ³ /mg	Class	TR	(mg/kg-dy) ⁻¹	(ug/m ³) ⁻¹	(mg/kg-dy) ⁻¹	mg/kg	THI	mg/kg-dy	mg/kg-dy	mg/kg-dy	mg/kg	mg/kg
Arsenic, Default	7440382	NA	Α	1.E-05	1.51E+01	4.30E-03	1.50E+00	2.08E+02	1	3.00E-04	1.50E-05	4.29E-06	1.33E+02	1.33E+02
Arsenic, RBA Adjusted	7440382	NA	A	1.E-05	1.51E+01	4.30E-03	2.03E-01	1.54E+03	1	2.22E-03	1.50E-05	4.29E-06	9.89E+02	9.89E+02

Data Input
Database look up values
Spreadsheet calculation

NA - Not Applicable, nonvolatile substance

	SF。	RfD _o
	(mg/kg-dy) ⁻¹	mg/kg-dy
Arsenic Default	1.50E+00	3.00E-04
Arsenic, RBA Adjusted	2.03E-01	2.22E-03

SF_o, RBA Adjusted = SF_o Default x RBA

 $RfD_{o},\,RBA$ Adjusted = RfD_{o} Default / RBA

RBA = 0.0135 or 13.5%

Table B-4 Type 5 Construction Worker Soil RRSs

Type 5 Risk Reduction Standards (RRSs) pose no significant risk on the basis of <u>SITE</u> <u>SPECIFIC EXPOSURE ASSUMPTIONS</u> for <u>NON-RESIDENTIAL PROPERTIES - TRESPASSER</u>									
RAGS Equation 6									
	Non-Residental Trespasser - Carcinogenio	c Effects							
C, ma/ka =	<u>TR x BW x AT_{car} x 365 days</u>	<u>/year</u>							
	¹⁹ EF x ED x [(SFo x 10-6 kg/mg x IRsoil) + (SFi x IRair x {1/VF + 1/PEF})]								
Parameter	Definition (units)	Default Value	Source						
C _{car} =	Concentration in soil (mg/kg)	Calculated	Not Applicable						
TR _{A/B}	IRIS Carcinogen Class A/B target excess lifetime cancer risk	1.E-05	HSRA Rules						
TR _C	IRIS Carcinogen Class C target excess lifetime cancer risk	1.E-05	HSRA Rules						
SF	Inhalation cancer slope factor (mg/kg-dy) ⁻¹	Chemical-Specific	RSLs						
- . [= IUR x 1,000 x BW 70 kg / IR _{air} 20 m ³ /dy								
IUR	Inhalation unit risk (ug/m ³) ⁻¹	Chemical-Specific	RSLs						
SFo	Oral cancer slope factor (mg/kg-dy) ⁻¹	Chemical-Specific	RSLs						
AT _{car-adult}	Adult averaging Time (yr)	70	HSRA Rules						
AT _{car-adolescent}	Adolescent averaging Time (yr)	70	HSRA Rules						

RAGS Equation 7 Non-Residental Trespasser - Noncarcinogenic Effects							
C _{noncar} mg/kg =	<u>THI x BW x ATnoncar x 365 day</u> ED x EF x [{(1/RfDo) x 10-6 kg/mg x IRsoil} + {(1/RfD	<u>s/year</u> i) x IRair x (1/VF	+ 1/PEF)}]				
Parameter	Definition (units)	Default Value	Source				
C _{noncar} =	Concentration in soil (mg/kg)	Calculated	Not Applicable				
THI	Target hazard index (none)	1	HSRA Rules				
RfD _o	Oral chronic reference dose (mg/kg-dy)	Chemical-Specific	RSLs				
RfD _i	Inhalation chronic reference dose (mg/kg-dy)	Chemical-Specific	RSLs				
RfC _i	Inhalation reference concentration (mg/m ³)	Chemical-Specific	RSLs				
AT _{noncar-adult}	Adult averaging Time (yr)	30	HSRA Rules				
AT _{noncar-adolescent}	Adolescent averaging Time (yr)	10	HSRA Rules				

Table B-5: Type 5 Trespasser Soil RRS Equations/Assumptions



Standard Assumptions							
BW _{adult}	Adult Body weight (kg)	70	HSRA Rules				
BW _{adolescent}	Adolscent body weight (kg)	45	Site-Specific				
EF	Exposure frequency (dy/yr)	100	Site-Specific				
ED _{adult}	Adult exposure duration (yr)	30	Site-Specific				
ED _{adolescent}	Adolescent exposure duration (yr)	10	Site-Specific				
IR _{air-adult}	Adult inhalation rate (m ³ /dy)	20	HSRA Rules				
IR _{air-adolescent}	Adolescent inhalation rate (m ³ /dy)	20	HSRA Rules				
IR _{soil-adult}	Adult soil ingestion rate (mg/dy)	100	Site-Specific				
IR _{soil-adolescent}	Adolescent soil ingestion rate (mg/dy)	200	Site-Specific				
PEF	Particulate emission factor (m ³ /kg)	4.63E+09	HSRA Rules				

	Soil-to-Air Volatilization Factor (VF)		
VF (m ³ /kg) =	<u>(LS x V x DH) x (π x α x T)</u> ¹ A x 2 x D _{ei} x E x K _{as} x 10 ⁻³ kg	/2 	
Parameter	Definition (units)	Default Value	Source
LS	Length of side of contaminated area (m)	45	HSRA Rules
V	Wind speed in mixing zone (m/s)	2.25	HSRA Rules
А	Area of contamination (cm ²)	2.03E+07	HSRA Rules
DH	Diffusion height (m)	2	HSRA Rules
α	$(D_{ei} \times E)/[E + (\rho_s \times (1-E)/K_{as})] (cm^2/s)$	Calculated	HSRA Rules
Т	Exposure Interval (s)	7.90E+08	HSRA Rules
ρ _s	Density of soil solids (g/cm ³)	2.65	HSRA Rules
D _{ei}	Effective diffusivity (cm ² /s)	D _i x E ^{0.33}	HSRA Rules
D _i	Molecular Diffusivity (cm²/s)	Chemical-specific	RSLs
E	Total soil porosity	0.35	HSRA Rules
K _{as}	Soil-air partition coefficient (g soil/cm ³ air)	(H/K _d) x 41	HSRA Rules
Н	Henry's Law Constant (atm-m ³ /mole)	Chemical-specific	RSLs
K _d	Soil-water partition coefficient (cm ³ /g)	K _{OC} x OC	HSRA Rules
K _{oc}	Organic carbon partition coefficient (cm ³ /g)	Chemical-specific	RSLs
OC	Soil Organic Carbon Content (none)	2.0E-02	HSRA Rules

HSRA Rules: Georgia Hazardous Response Act Rules, 391-3-19, Appendix III, Media Target Concentrations and Standard Exposure Assumptions.

http://rules.sos.state.ga.us/docs/391/3/19/Appendix%20I-IV.pdf

RAGS: Risk Assessment Guidance for Superfund, Volume I - Human Health Evaluation Manual (Part B, Chapter 3, Development of Risk-Based Preliminary Remediation Goals), U.S. Environmental Protection Agency, December 1991. http://www.epa.gov/oswer/riskassessment/ragsb/pdf/chapt3.pdf

> RSLs: U.S. Environmental Protection Agency Regional Screening Levels, November 2017 https://www.epa.gov/risk/regional-screening-levels-rsls-generic-tables-november-2017

Table B-5: Type 5 Trespasser Soil RRS Equations/Assumptions



North Berkeley Lake Road Site (HSI 10844) Fire Station 19 Duluth, Gwinnett County, Georgia

		Carcinogenic Effects ¹						Noncarcinogenic Effects ¹								
																Type 5
		VF	Car.		SFi	IUR	\mathbf{SF}_{o}	C _{car-adult}	C _{car-adolescent}		RfD_{o}	RfC _i	RfD _i	C _{noncar-adult}	C _{noncar-adolescent}	RRS
Substance	CAS No.	m ³ /mg	Class	TR	(mg/kg-dy) ⁻¹	(ug/m ³) ⁻¹	(mg/kg-dy) ⁻¹	mg/kg	mg/kg	THI	mg/kg-dy	mg/kg-dy	mg/kg-dy	mg/kg	mg/kg	mg/kg
Arsenic, Default	7440382	NA	А	1.E-05	1.5E+01	4.3E-03	1.5E+00	4.8E+02	4.6E+02	1	3.0E-04	1.5E-05	4.3E-06	9.2E+03	3.0E+03	4.6E+02
Arsenic, RBA Adjusted	7440382	NA	A	1.E-05	1.5E+01	4.3E-03	2.0E-01	3.5E+03	3.4E+03	1	2.2E-03	1.5E-05	4.3E-06	6.8E+04	2.2E+04	3.4E+03

1 - Per EPD's recommendation, a conversion factor of 0.0833 has been applied to account for an exposure time of 2 hours per day.

Data Input
Database look up values
Spreadsheet calculation

NA - Not Applicable, nonvolatile substance

SFo	RfD_{o}
(mg/kg-dy) ⁻¹	mg/kg-dy
1.50E+00	3.00E-04
2.03E-01	2.22E-03
	SF o (mg/kg-dy) ⁻¹ 1.50E+00 2.03E-01

SF_o, RBA Adjusted = SF_o Default x RBA RfD_o, RBA Adjusted = RfD_o Default / RBA RBA = 0.0135 or 13.5%

Table B-6Type 5 Trespasser Soil RRSs Calculations

CDM Smith



